A background image showing several micro-chip chromatography components, including a chip with a grid pattern and a chip with a single channel, all in a teal color scheme.

# The micro-Chip Chromatography Company

PharmaFluidics Team  
Please visit [www.pharmafluidics.com](http://www.pharmafluidics.com)



MD Scientific is an authorized distributor in Denmark  
[www.md-scientific.dk](http://www.md-scientific.dk) - +45 7027 8565

---

# PharmaFluidics - Company Profile

Spin-off of Vrije Universiteit Brussel

Located near Ghent, Belgium in “Flanders’ Life Sciences valley”

Industry:

- ALDA : Global Analytical Lab, Life Sciences & Diagnostics Instrumentation Industry
- **HPLC/MS : High Performance Liquid Chromatography / Mass Spectroscopy Segment**

Key expertise

- Design, lithographic production, surface treatment of micro-Chip based HPLC columns

**Product:  $\mu$ PAC™ - an new generation of HPLC columns**

USP:

Detect more molecules, with higher sensitivity, in tiny, complex biological samples

Market:

Premium segment “ultra-high resolution” LC/MS market

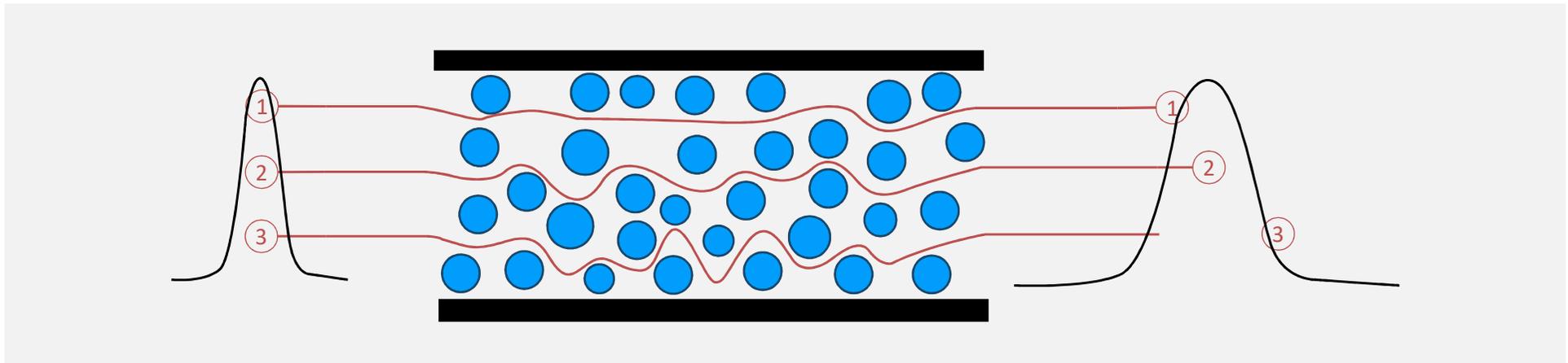
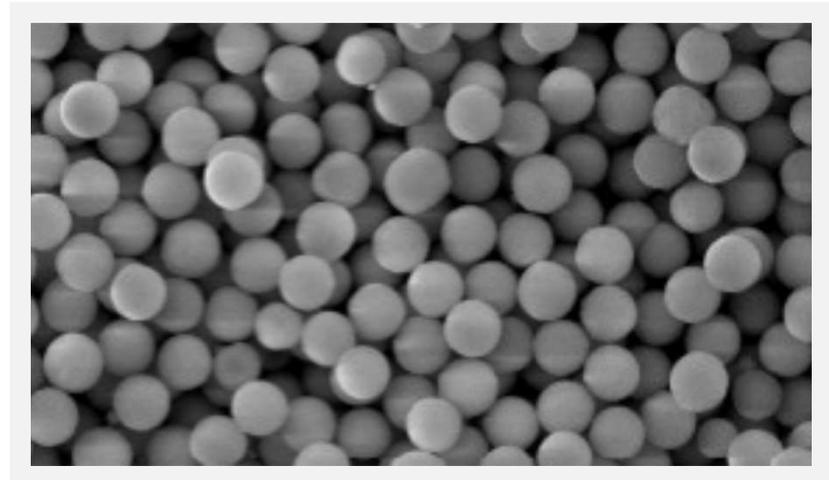
Sales:

- Presented to market at specialist trade shows in the USA and Europe since spring 2017
- Priced in June 2017
- Sold since September 2017 for Proteomics, Metabolomics and Lipidomics profiling, to Bio-Pharma customers with bio-marker, diagnostics and drug research & development applications

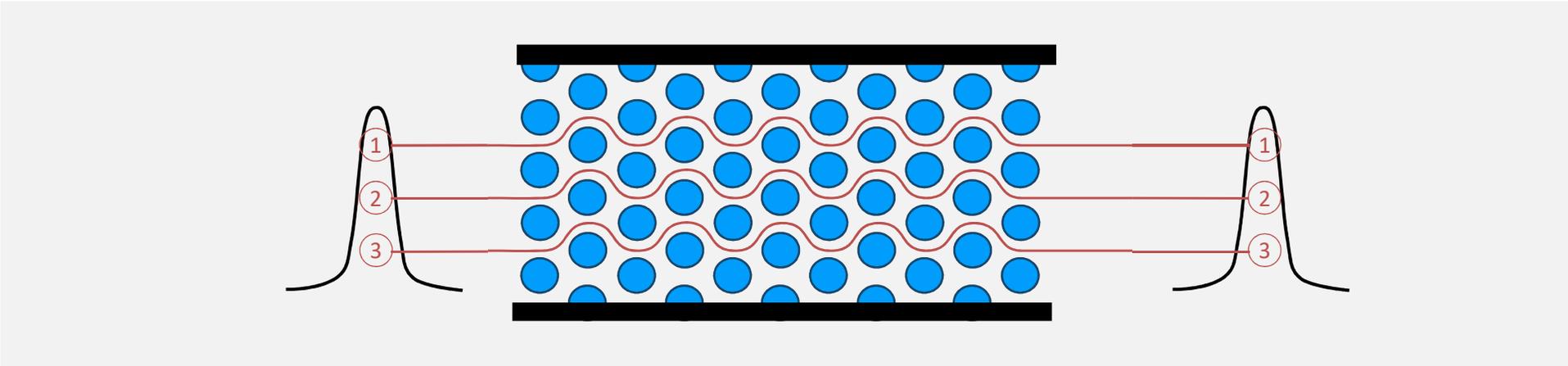
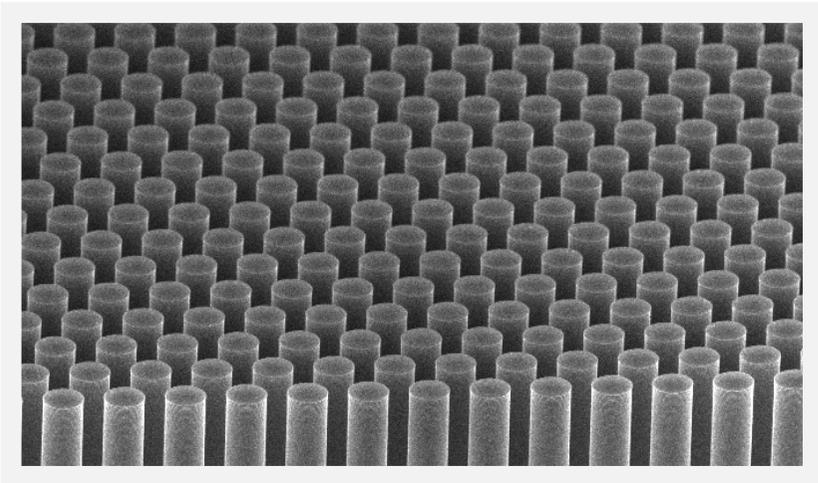
# The Micro Pillar Array Column ( $\mu$ PAC™)

---

# Traditional HPLC columns

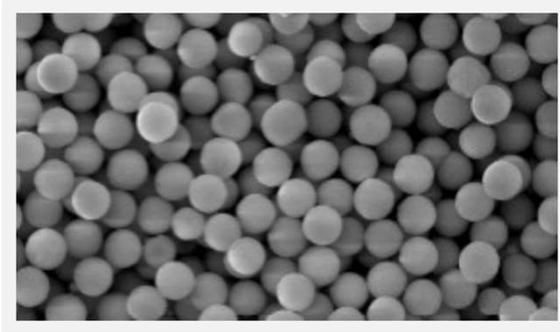


# PharmaFluidics' game-changing technology

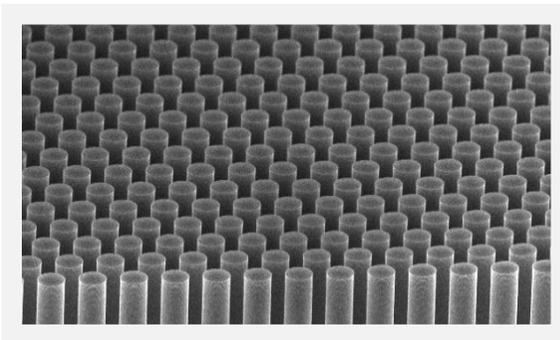
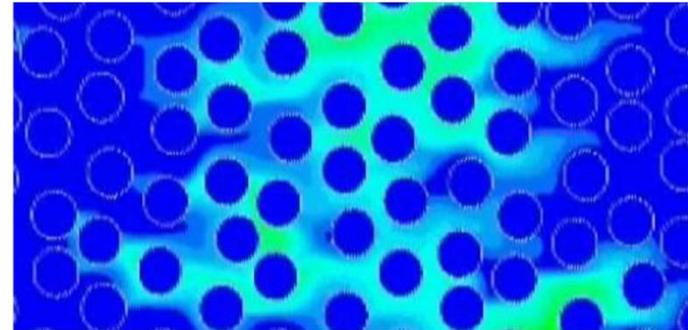


# Unprecedented separation performance

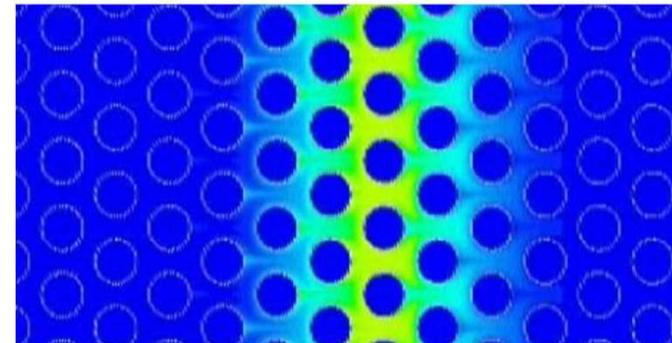
The benefit of Order versus Disorder



Disorder - Packed Bed

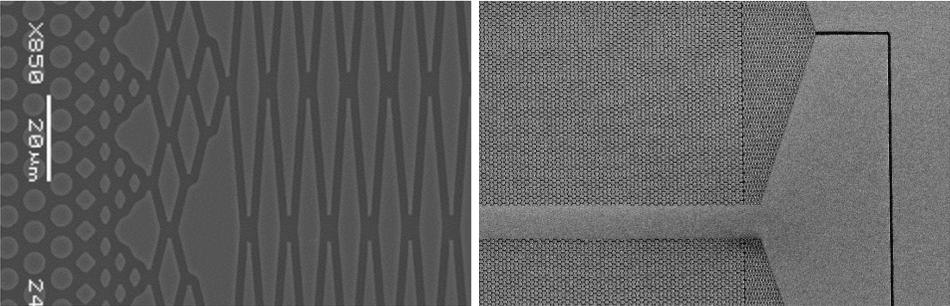


Order – Pillar Array



# Unprecedented separation performance

μPAC™ - Pillar Array Columns

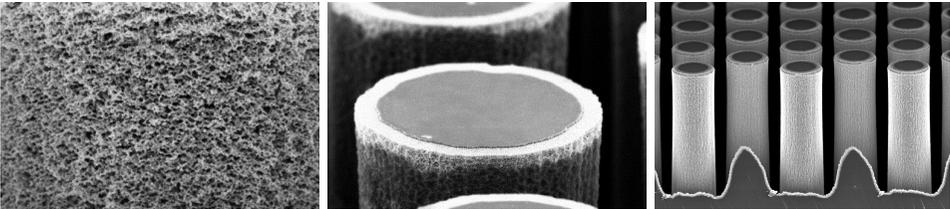


Flow distribution for perfect plug conservation in inlets and turns



*200 cm column*

*50 cm column*



Stationary Phase controlled by design

---

# Unprecedented separation performance

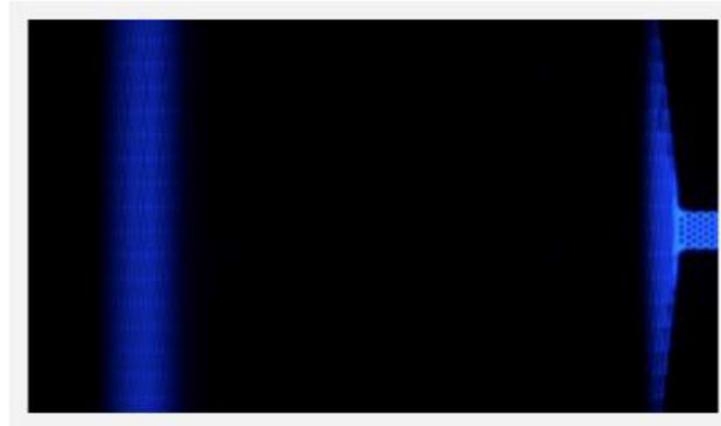
μPAC™ - Pillar Array Columns



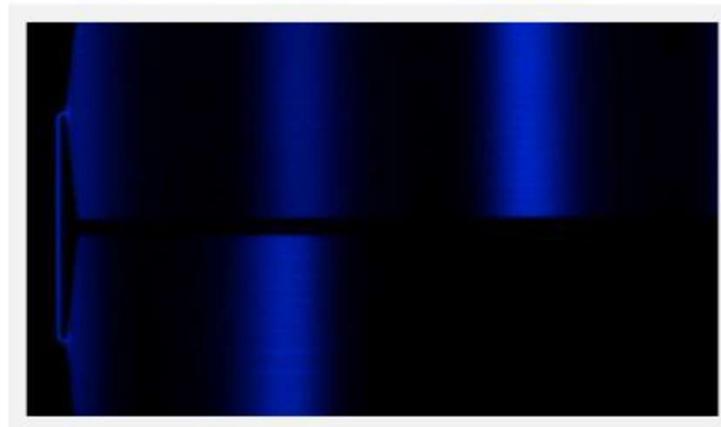
---

## Flow distribution

Unprecedented separation bed length on a small footprint, without additional peak dispersion



From narrow in- and outlet to wider separation channels



Connecting parallel Separation channels

# μPAC™ - C18 – 200 cm/50 cm

**micro-Chip  
Liquid Chromatography Cartridges  
for use with  
Any 3<sup>rd</sup> party HPLC/MS equipment**

**Highly reliable and robust nano-LC  
solution with:**

- The highest resolution
- The best data productivity
- State-of-the-art repeatability

**Allowing unique modes of operation**

- Pressure at optimal flow rate of 300 nl/min ~ 100 bar (200cm μPAC™) or ~ 40 bar (50cm μPAC™)
- Compliant up to 350 bar (5,000 psi) allowing a maximum flow rate of ~ 1 μl/min (200cm) or ~ 2.5 μl/min (50cm)





- ✓ Length of the column: 200cm
- ✓ Coating: C18-End capped
- ✓ Flow: max. 1 $\mu$ l/min (at 50°C)
- ✓ Max. pressure allowed: 320 bar/4641psi
  - ✓ At 300 nl/min (30°C): 90 – 120 bar/1305,34 – 1740,45psi
  - ✓ At 1 $\mu$ l/min (50°C): 80 – 320 bars/ 1160,30 – 4641,21psi

- ✓ Pillar distance: 2,5 $\mu$ m
- ✓ Pillar diameter: 5 $\mu$ m
- ✓ Pore size: 10 – 30nm
- ✓ Inlet union: 1/32" – 1/32"
- ✓ Outlet union: 1/32" – 360 OD

- ✓ Void volume: 9 $\mu$ l
- ✓ Outer dimensions:
  - L: 11cm/4.33inch
  - W: 3cm/1.18inch
  - H: 0,6cm/0,23inch
- ✓ Sample volume: >4nl

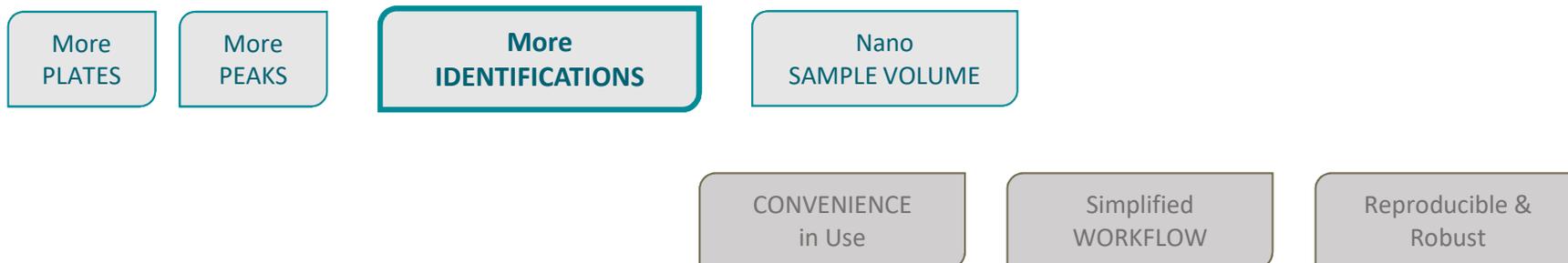


# Unprecedented separation performance

See more from less sample

Performance	<b>μPAC™ Chip Cartridge</b>	State-of-the-art HPLC	Standard HPLC
Plate number	400.000	250.000/m	150.000/m
Peak capacity	Towards 1.200	700	300

## Product USP



# Target markets and applications

## Biomarker discovery

Launch segment: metabolomics / lipidomics

### Challenges

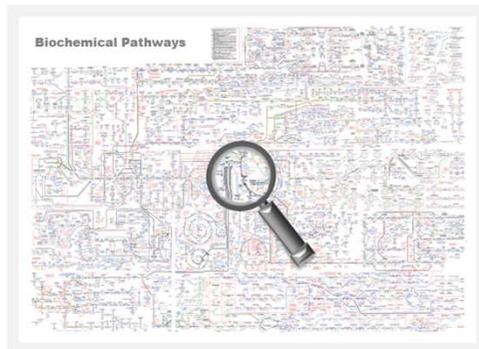
- Rare molecules in complex mixtures
- Tiny biopsies

## Bio-pharmaceuticals & Bio-similars development

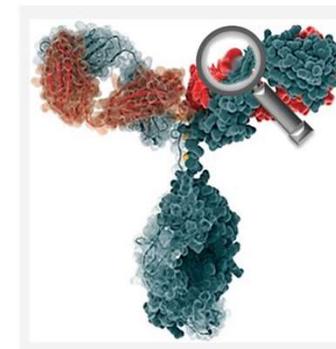
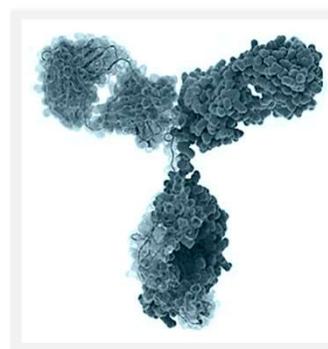
### Challenges

- Tiny differences in deamidation & glycosylation patterns of proteins or mono-clonal-antibodies
- Analytics supported FDA/EMA approvals

### Trace amounts in complex samples



### Subtle modifications

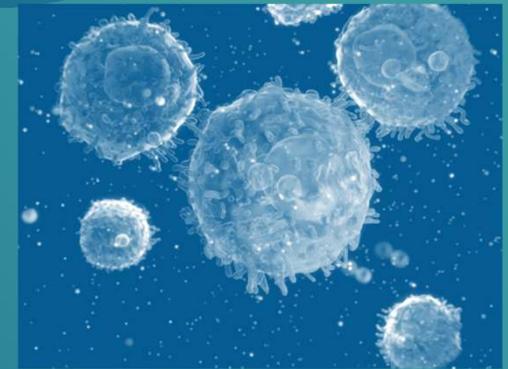


# Data and results

- $\mu$ PAC™-MS Proteomics Platform
  - $\mu$ PAC™-MS Lipidomics Platform
  - $\mu$ PAC™-MS Metabolomics Platform
  - $\mu$ PAC™-MS Biopharmaceuticals
-

# Data and results

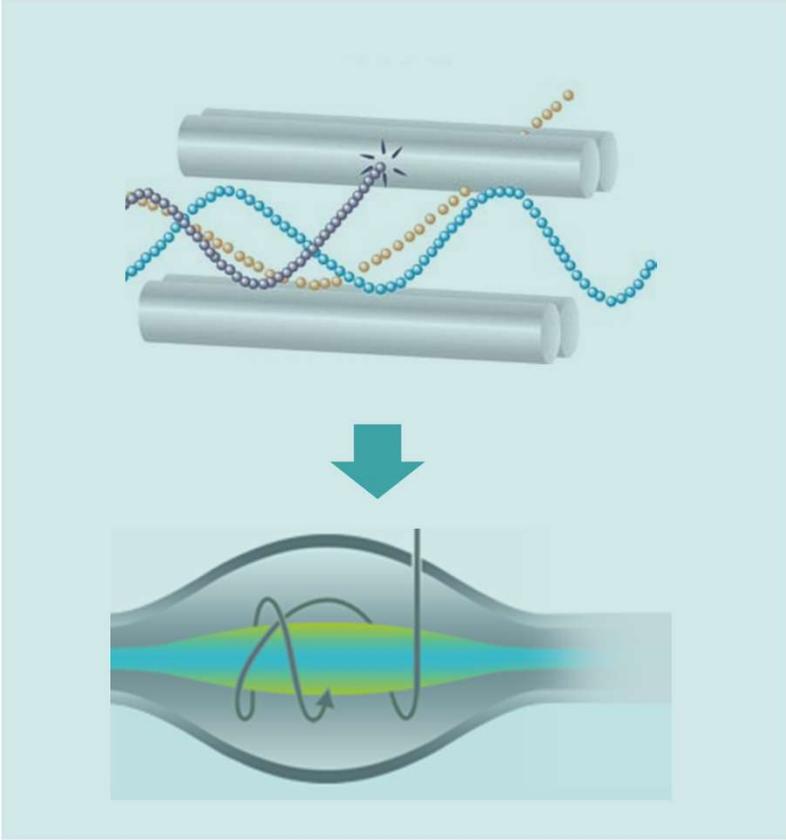
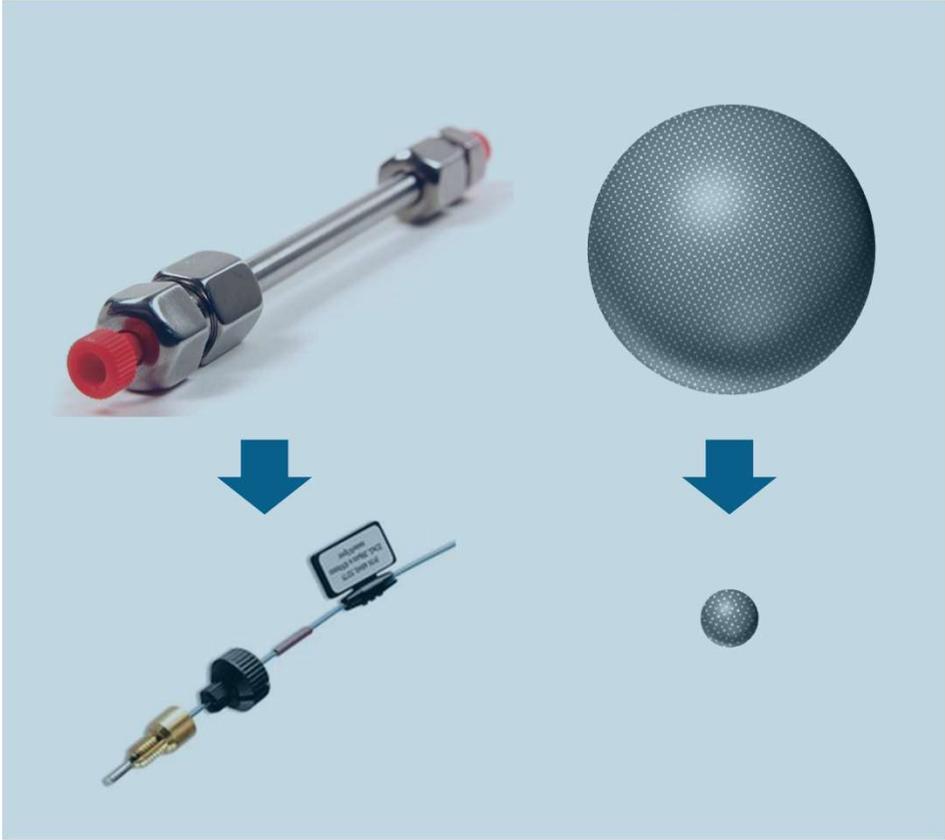
- **μPAC™-MS Proteomics Platform**
  - μPAC™-MS Lipidomics Platform
  - μPAC™-MS Metabolomics Platform
  - μPAC™-MS Biopharmaceuticals
- 



# State of the art in bottom-up proteomics

## LC

## MS



Sensitivity ↑

Efficiency ↑

Resolution & Sensitivity ↑

# State of the art in bottom-up proteomics



## LC systems

Flow rate: 50-1,500 nl/min  
Pressure:  $\leq 1,200$  bar  
(17,000 psi)



## LC columns

ID:  $75\ \mu\text{m}$   
Packing:  $\leq 2\ \mu\text{m}$  C18 silica  
Length:  $\geq 50$  cm



## Mass analyzers

Orbitrap-quadrupole hybrid  
...

Identification of over 4,000 proteins in a single 90-140 min gradient run

---

## Traditional HPLC column limitations

- High backpressure observed for long ( $\geq 50$  cm) LC columns
  - loss of data, upper pressure limit exceeded
  - wear & tear of LC pump system and fluidics
- Column packing is a delicate process
  - poor column-to-column reproducibility

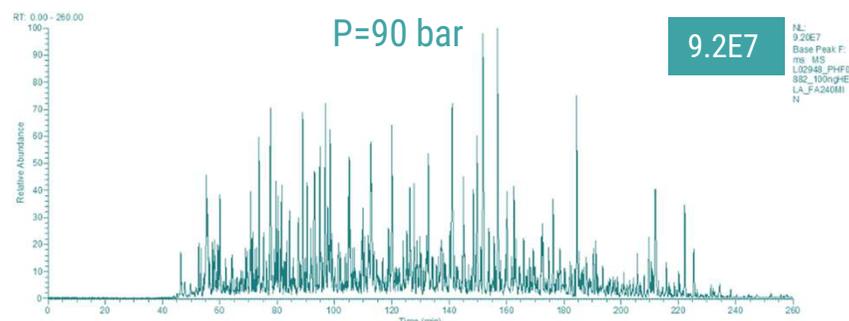
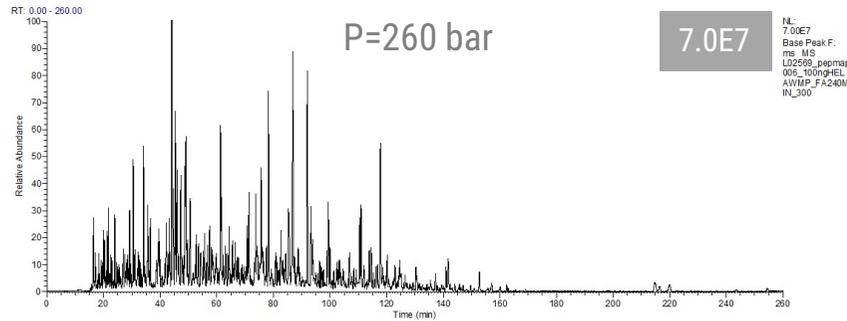
### How to increase column performance even further?

- Reduction of particle diameter → Increase in backpressure
- Increase in column length → Increase in backpressure

# μPAC™-MS Proteomics Platform

0.75 mm x 15 cm packed bed C18 (2 μm)	
# unique PEPTIDE identifications	# PROTEIN identifications
<b>6196</b>	<b>1219</b>

200 cm μPAC™ C18	
# unique PEPTIDE identifications	# PROTEIN identifications
<b>10560</b> (GROUND)	<b>1736</b> (GROUND)



## Conclusion

μPAC™ vs packed bed = **70%** increase in peptide ID's, **42%** increase in protein ID's

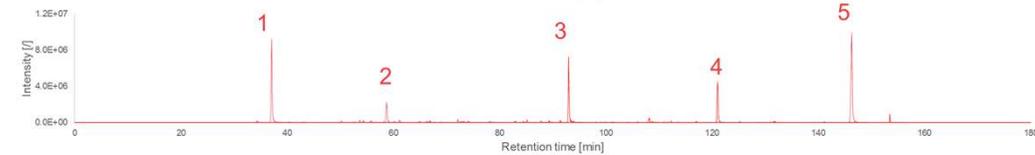
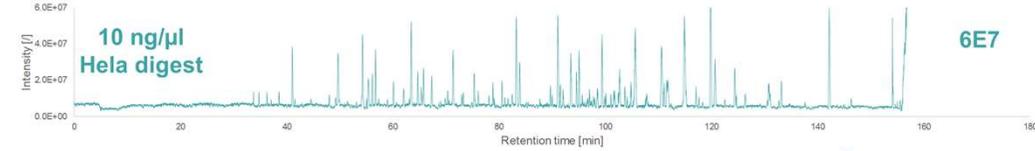
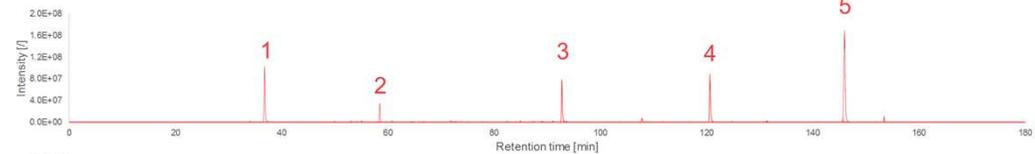
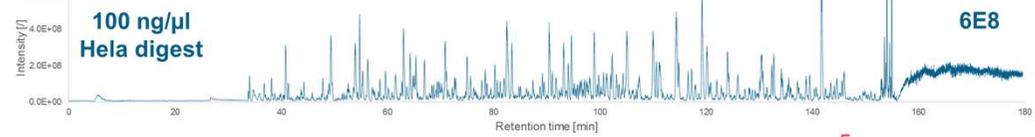
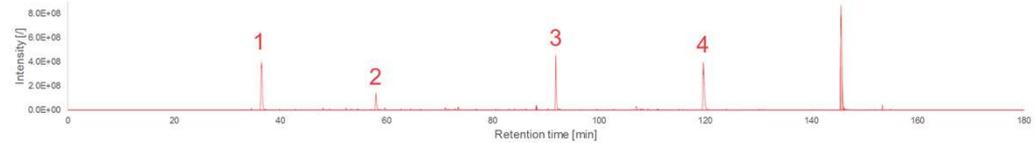
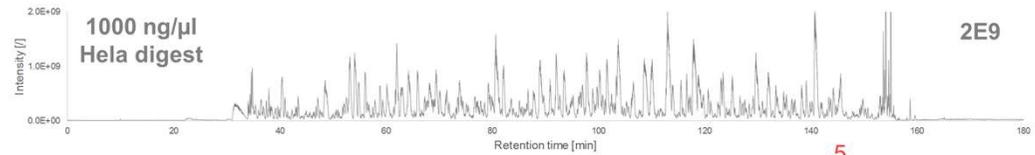
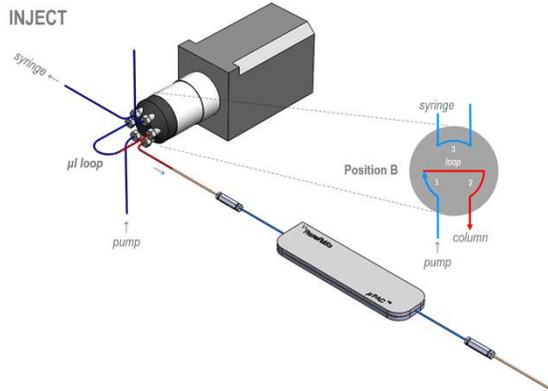
Obtained with LTQ Orbitrap

### Experimental conditions

Column(s): 200 cm μPAC™ C18  
15 cm Pepmap C18  
Flow: 0.3 μL/min  
Mobile Phase A: 0.1% FA  
Mobile Phase B: 0.1% FA in 80% ACN  
Gradient: 1-50% B in 240 min  
Temp: 30°C  
Inj vol: 1 μL  
Sample: 500 ng/μl HeLa cell digest  
Detection: Thermo LTQ Orbitrap XL MS

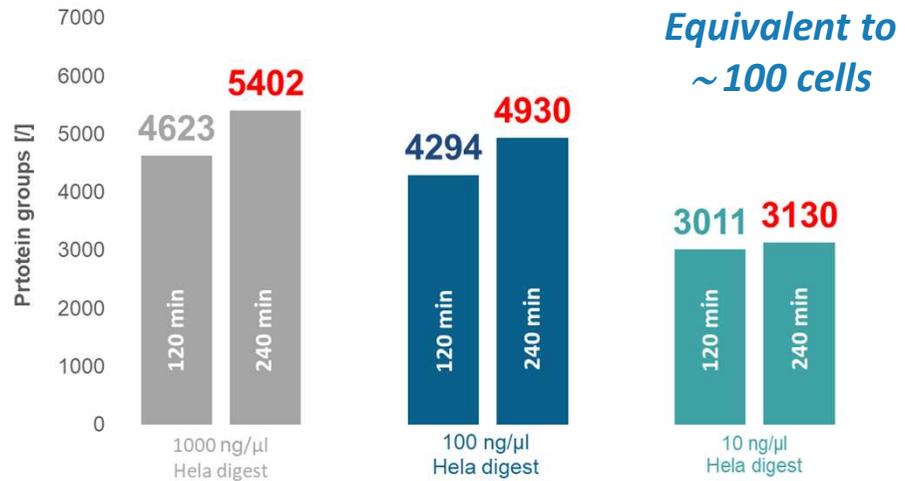
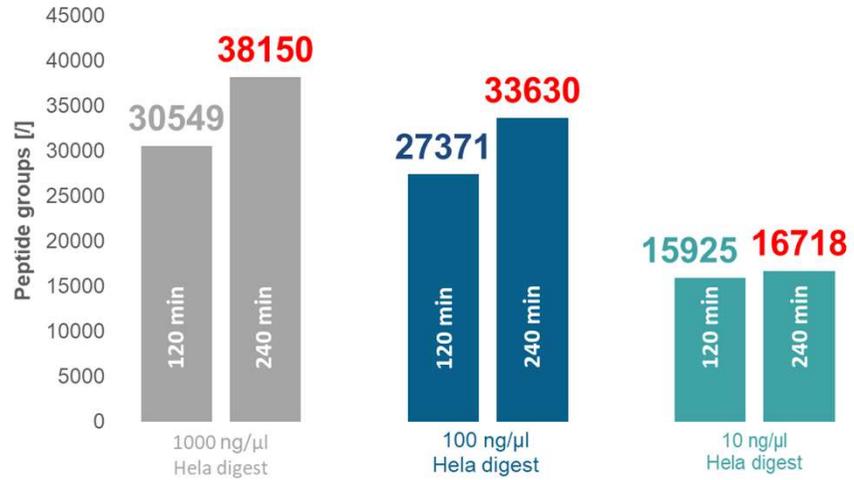
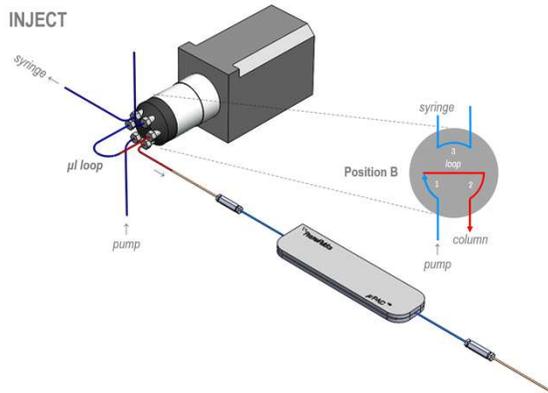
# μPAC™-MS Proteomics Platform

Hela cell digest – 200 cm μPAC™ connected to Orbitrap™ Fusion™ Lumos™ Tribrid™



# μPAC™-MS Proteomics Platform

Hela cell digest – 200 cm μPAC™ connected to Orbitrap™ Fusion™ Lumos™ Tribrid™

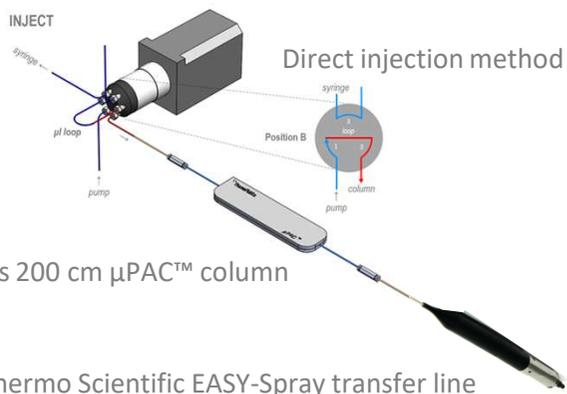


# μPAC™ Flexibility



## Experimental conditions

**Column:** 200 cm μPAC™ C18  
**Flow:** 1 μL/min  
**A:** 0.1% FA  
**B:** 0.1% FA in 80% ACN  
**Gradient:** 1-50% B in 30-60-90 min  
**Temp:** 50°C  
**Inj vol:** 2-0.1 μL  
**Sample:** 1000-500-250-100-50-10-2 ng/μl HeLa cell digest  
**Detection:** Thermo Orbitrap Q Exactive HFx



PharmaFluidics 200 cm μPAC™ column

Thermo Scientific EASY-Spray transfer line

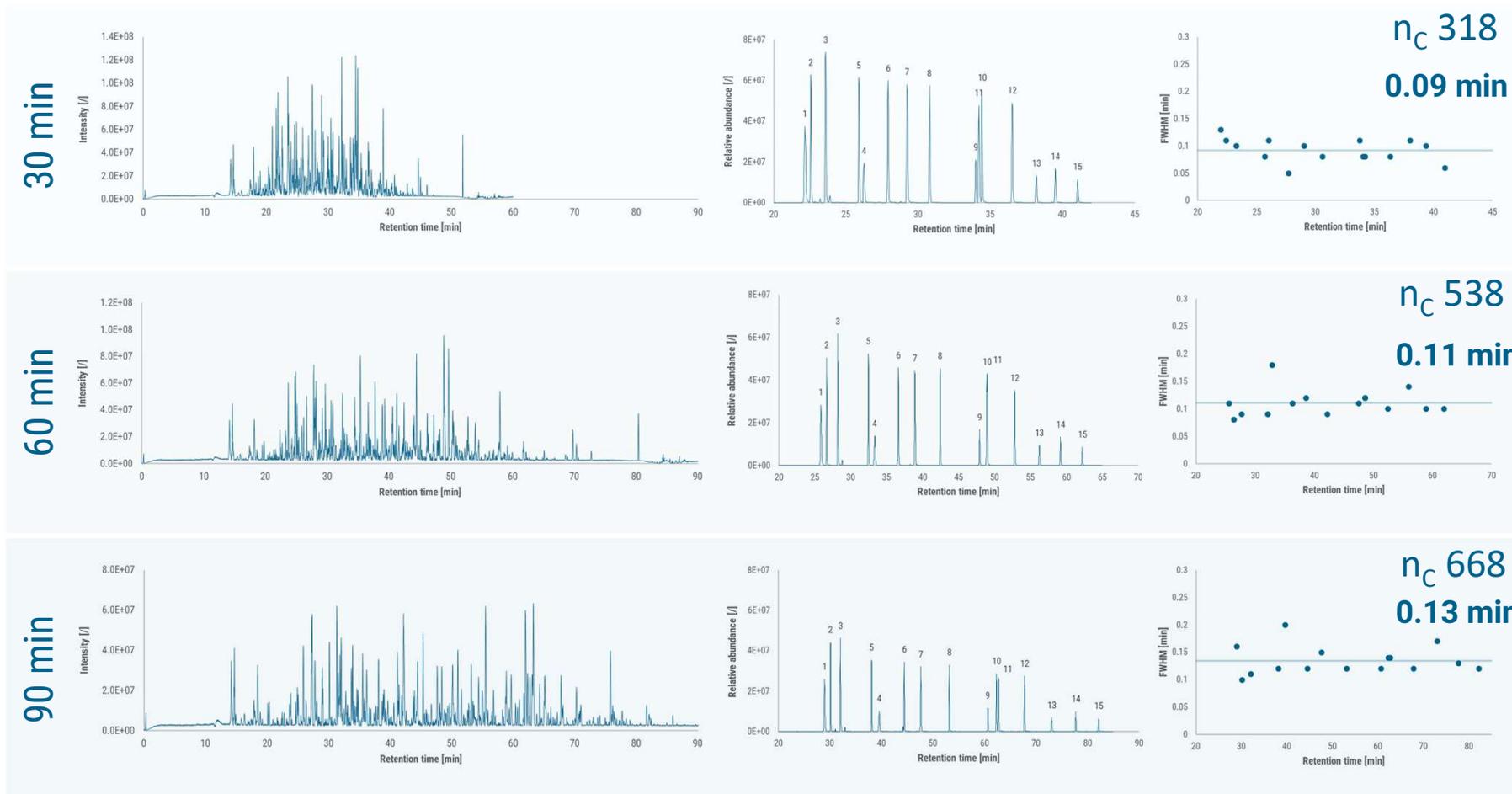
- **EASY-spray transfer line as ESI emitter**
- A dilution series of **HeLa cell tryptic digest** (1000 - 2 ng) separated using **30, 60 and 90 min** solvent gradient times.
- All samples spiked with an **internal peptide standard** (100 fmol/μl Pierce™ retention time calibration mixture) to evaluate the **chromatographic metrics** throughout the entire experiment.

# μPAC™ Flexibility

BPC 50 ng HeLa digest

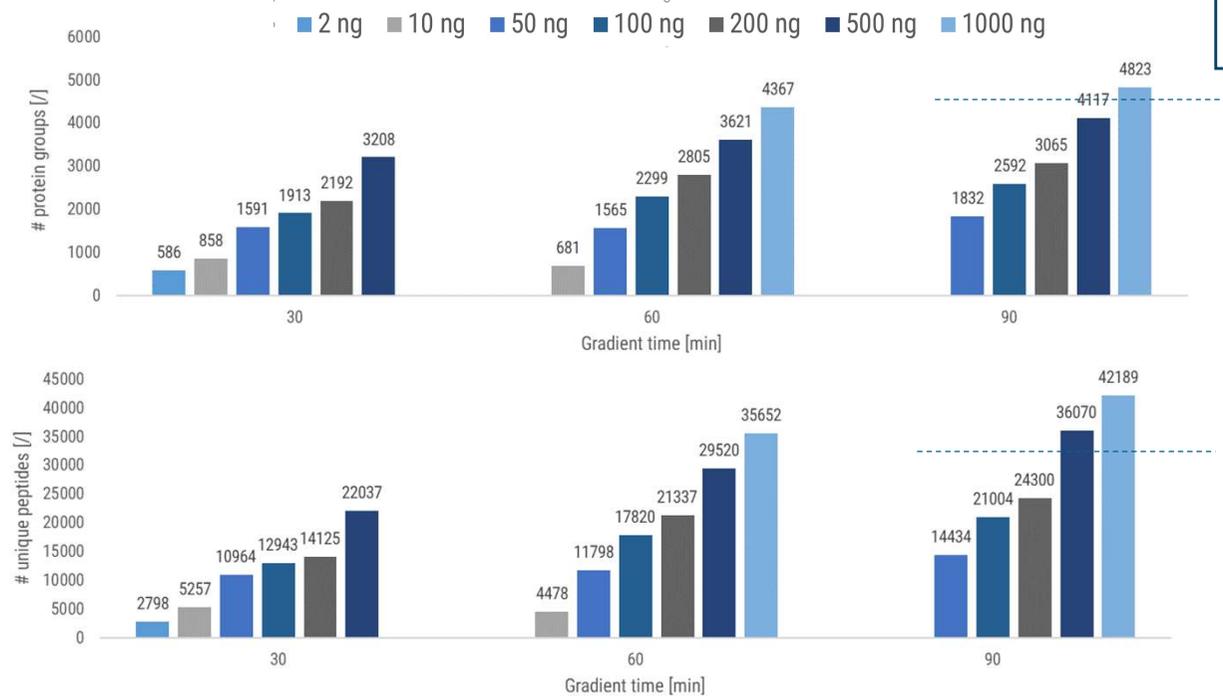
EIC 50 fmol PRTC

FWHM 50 fmol PRTC



# μPAC™ Flexibility

## Identifications HeLa cell digest separation



### Conclusions

- Retention time variation of 0.5% CV observed for 15 QC peptides (50-200 fmol/μl)
- Number of identifications with a 90 min gradient 'capillary' flow similar to a 120 min 'nano' flow run for routine sample amounts (1 μg)

# Performance test 50cm $\mu$ PAC

---

# μPAC™ 50cm - Benchmarking



## Experimental conditions

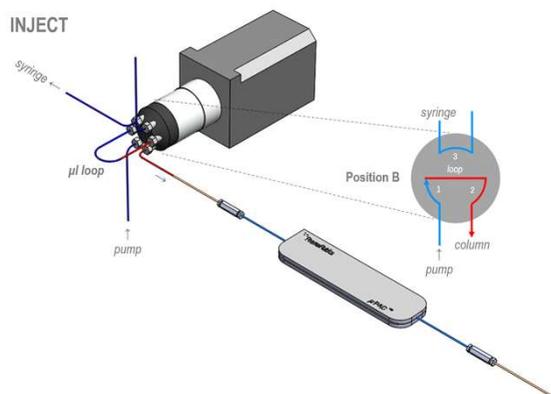
### Chromatography

Instrument: Thermo Scientific™ Ultimate 3000 RSLC nano  
Injection: 1 μL, user defined injection  
Column temp: 50°C  
Solvent A: 0.1% FA  
Solvent B: 0.1% FA in 80% ACN  
Gradient: 1-45% B in 30-60-90 min

### Mass Spectrometry

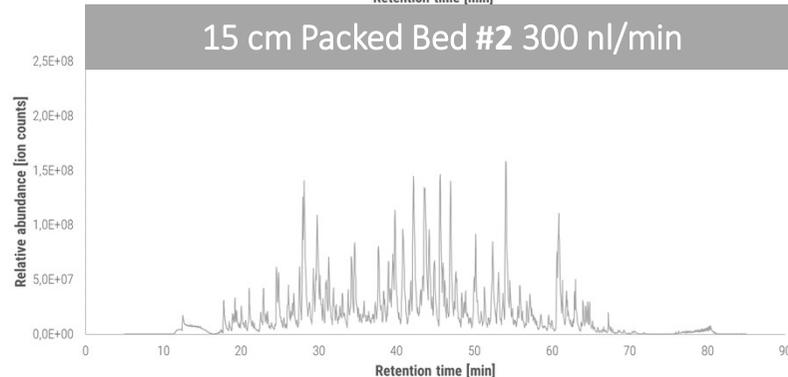
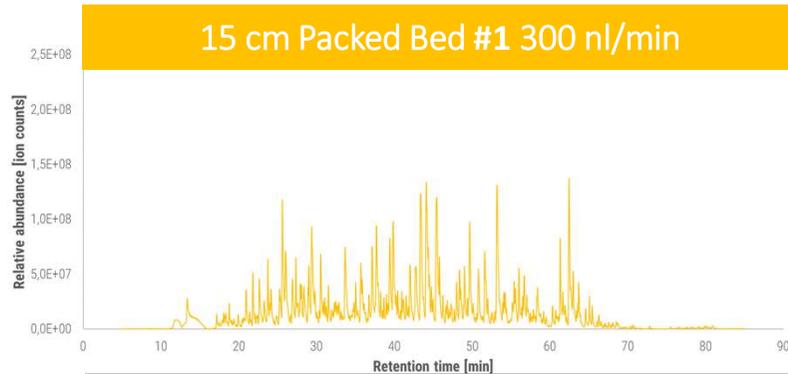
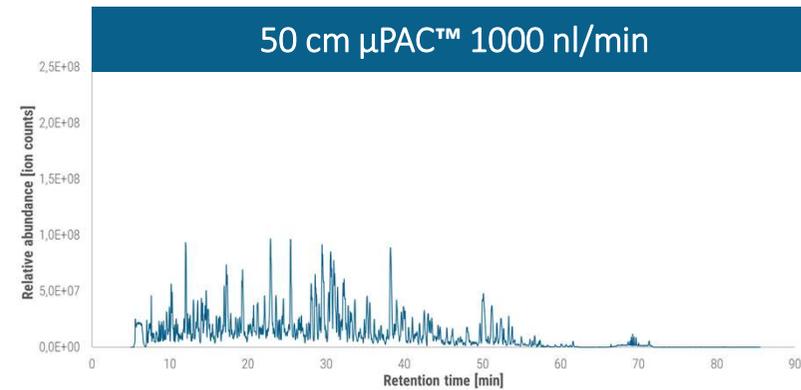
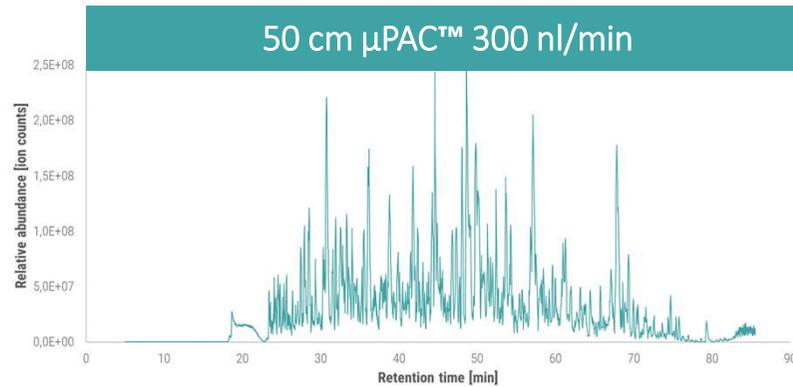
Instrument: Thermo Scientific™ LTQ Orbitrap Elite (DDA)  
Resolution: 60,000 (MS1); 15,000 (MS2)  
Mass range: 300-2000 m/z  
Max IT: 80 ms

Direct injection method



- Benchmarking μPAC™ 50cm column against conventional packed bed column (2 μm particles; 15cm x 75 μm)
- 500 ng **HeLa cell tryptic digest** was separated using **30, 60 and 90 min** solvent gradient times.
- All measurements were performed in triplicate

# 50 cm $\mu$ PAC™ - Benchmarking in Bottom-up Proteomics



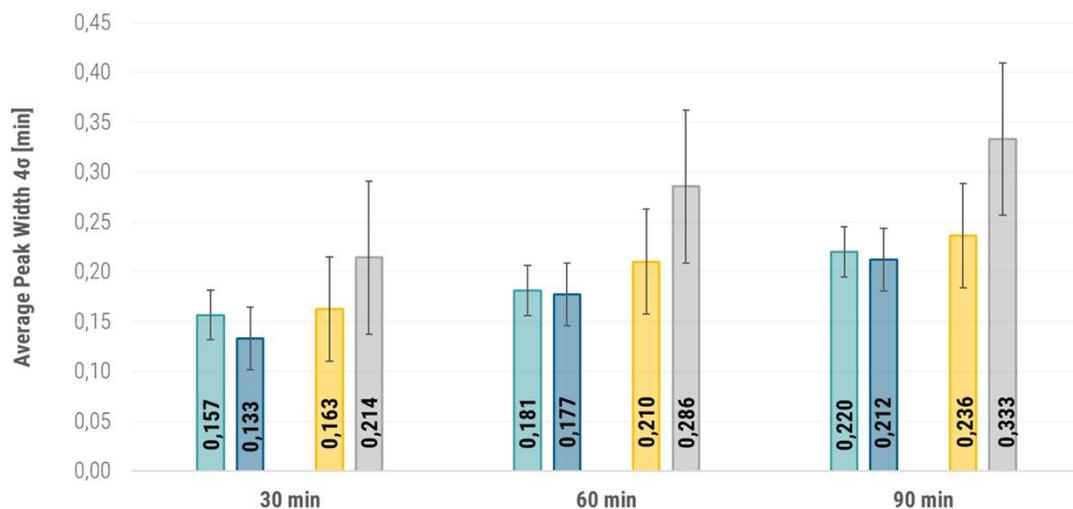
## Instrumental Set-up

Thermo Scientific™ Ultimate 3000 RSLC nano  
Thermo Scientific™ LTQ Orbitrap Elite™ (DDA)

1  $\mu$ L direct injection  
500 ng commercial HeLa digest + 50 fmol PRTC mix  
Gradient: 1-45% B  
A: H<sub>2</sub>O + 0.1% FA;  
B: 80% ACN + 0.1% FA

50cm  $\mu$ PAC™ column  
15 cm Packed Bed #1: (150 x 0.075 mm, 2.0  $\mu$ m particles)  
15 cm Packed Bed #2: (150 x 0.075 mm, 1.7  $\mu$ m particles)

# 50 cm $\mu$ PAC™ - Benchmarking in Bottom-up Proteomics

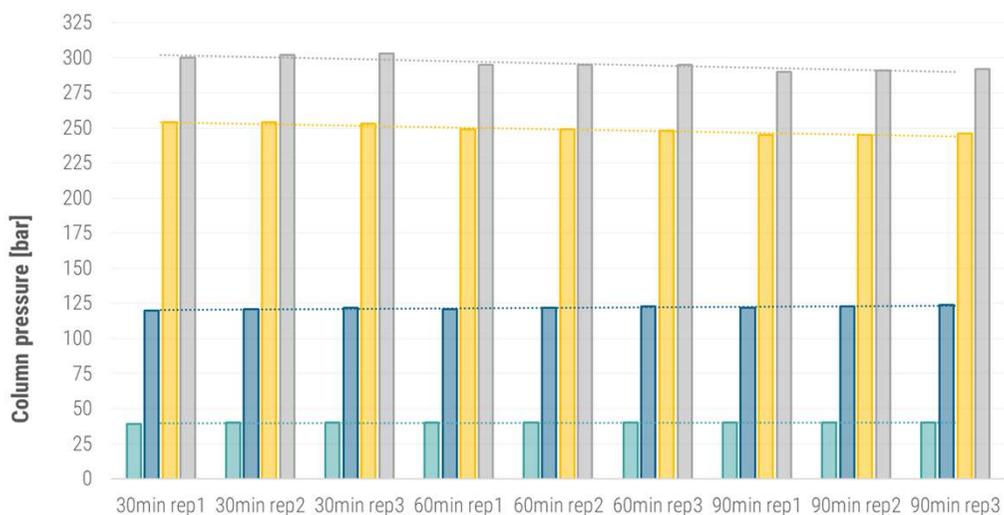


### Peptide Peak Width

30 min gradient: 0.13 - 0.16 min  
 60 min gradient: 0.17 - 0.18 min  
 90 min gradient: 0.21 - 0.22 min

→ 4 - 60% reduction

50 cm $\mu$ PAC™	300 nl/min
50 cm $\mu$ PAC™	1000 nl/min
15 cm Packed Bed #1	300 nl/min
15 cm Packed Bed #2	300 nl/min

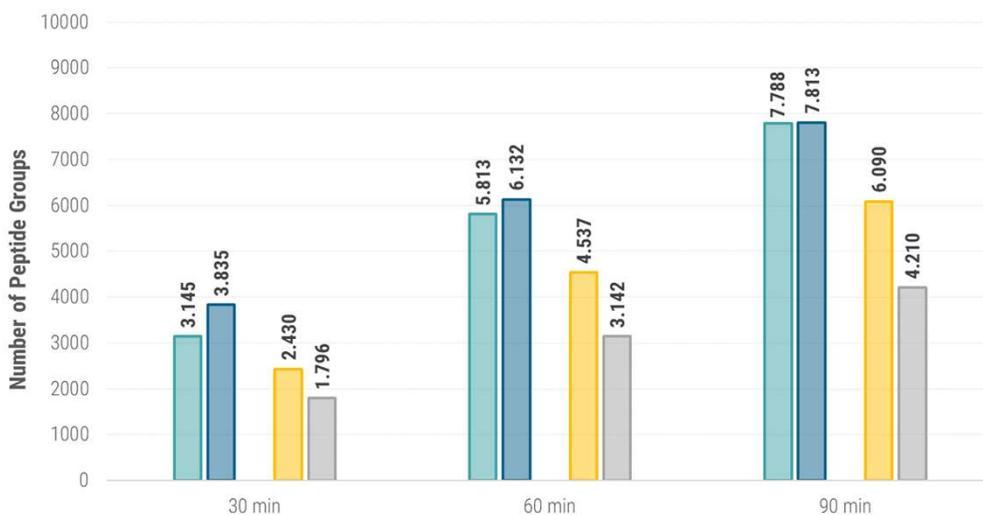
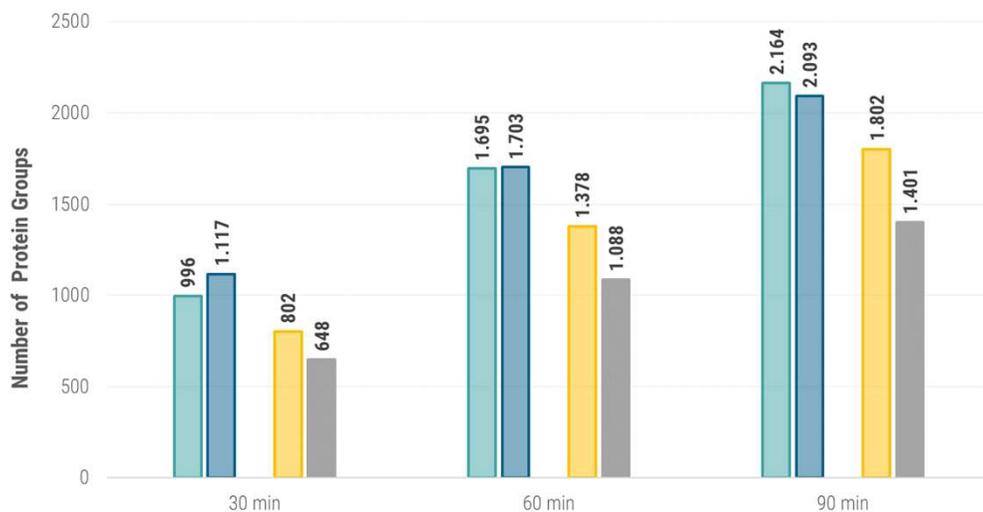


### Column Backpressure

300 nl/min: 40 bar  
 1000 nl/min: 120 bar

→ 6-8 times lower

# 50 cm $\mu$ PAC™ - Benchmarking in Bottom-up Proteomics



## Peptides

300 nl/min → 28 - 75% increase

1000 nl/min → 28 - 114% increase

50 cm  $\mu$ PAC™ 300 nl/min

50 cm  $\mu$ PAC™ 1000 nl/min

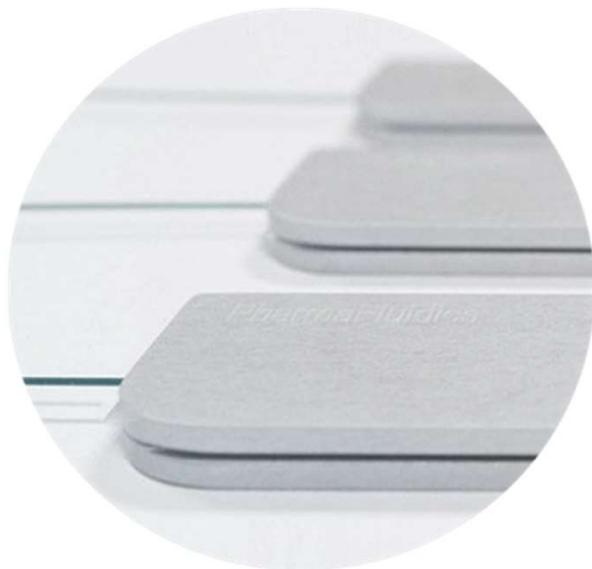
15 cm Packed Bed #1 300 nl/min

15 cm Packed Bed #2 300 nl/min

## Protein Groups

300 nl/min → 20 - 56% increase

1000 nl/min → 16 - 72% increase



### 50 cm μPAC™

- 3 μl column volume
- 0.1 – 2 μl/min
- 30 min – 2h analysis time

Increased Throughput



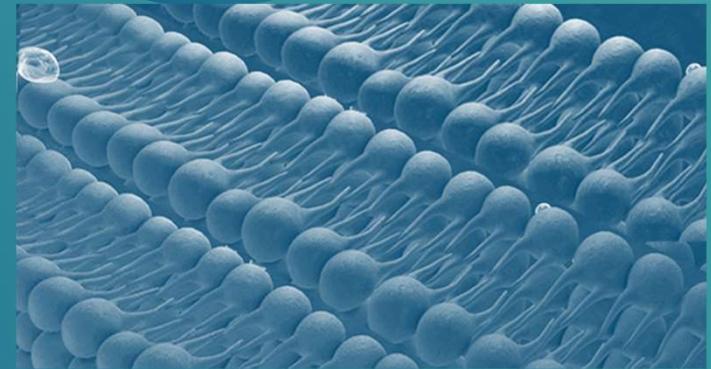
### 200 cm μPAC™

- 9 μl column volume
- 0.1 – 1 μl/min
- 2h – 8h analysis time

Unique Proteome Coverage

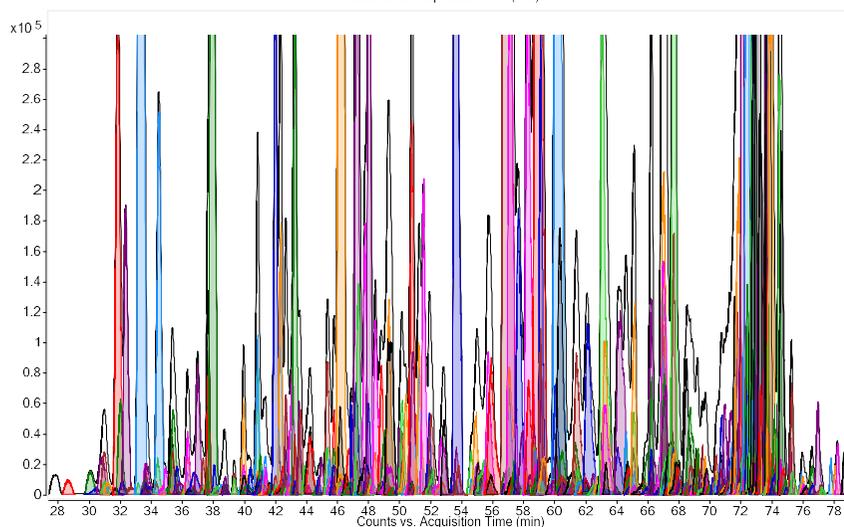
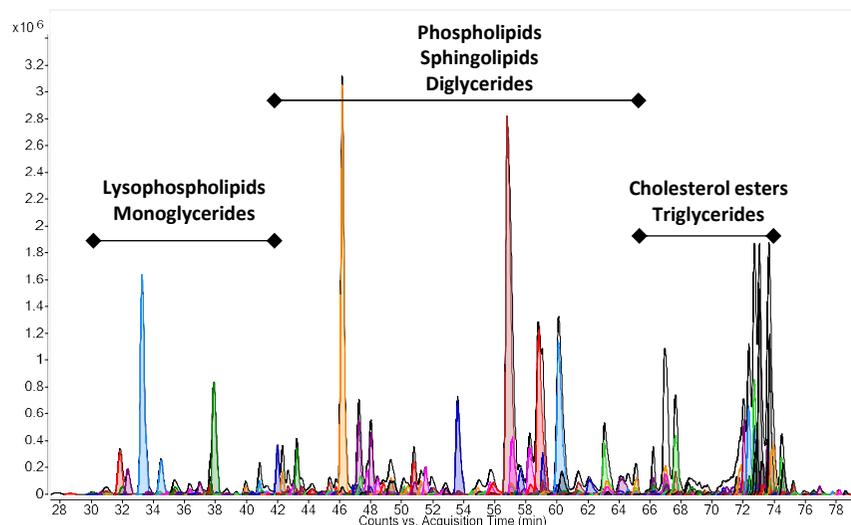
# Data and results

- $\mu$ PAC™-MS Proteomics Platform
  - **$\mu$ PAC™-MS Lipidomics Platform**
  - $\mu$ PAC™-MS Metabolomics Platform
  - $\mu$ PAC™-MS Biopharmaceuticals
- 



# μPAC™-MS Lipidomics Platform

## Separation of human blood plasma lipid extract



Enormous sample complexity is revealed, illustrative for μPAC-MS resolving power!

### INTER-class lipid separation

- All major lipid classes are detected
- Location indicated on the chromatogram

### INTRA-class lipid separation

- Number of carbons
- Degree of saturation in fatty acid side chains
- Fatty acid side chain position & composition

### Experimental conditions

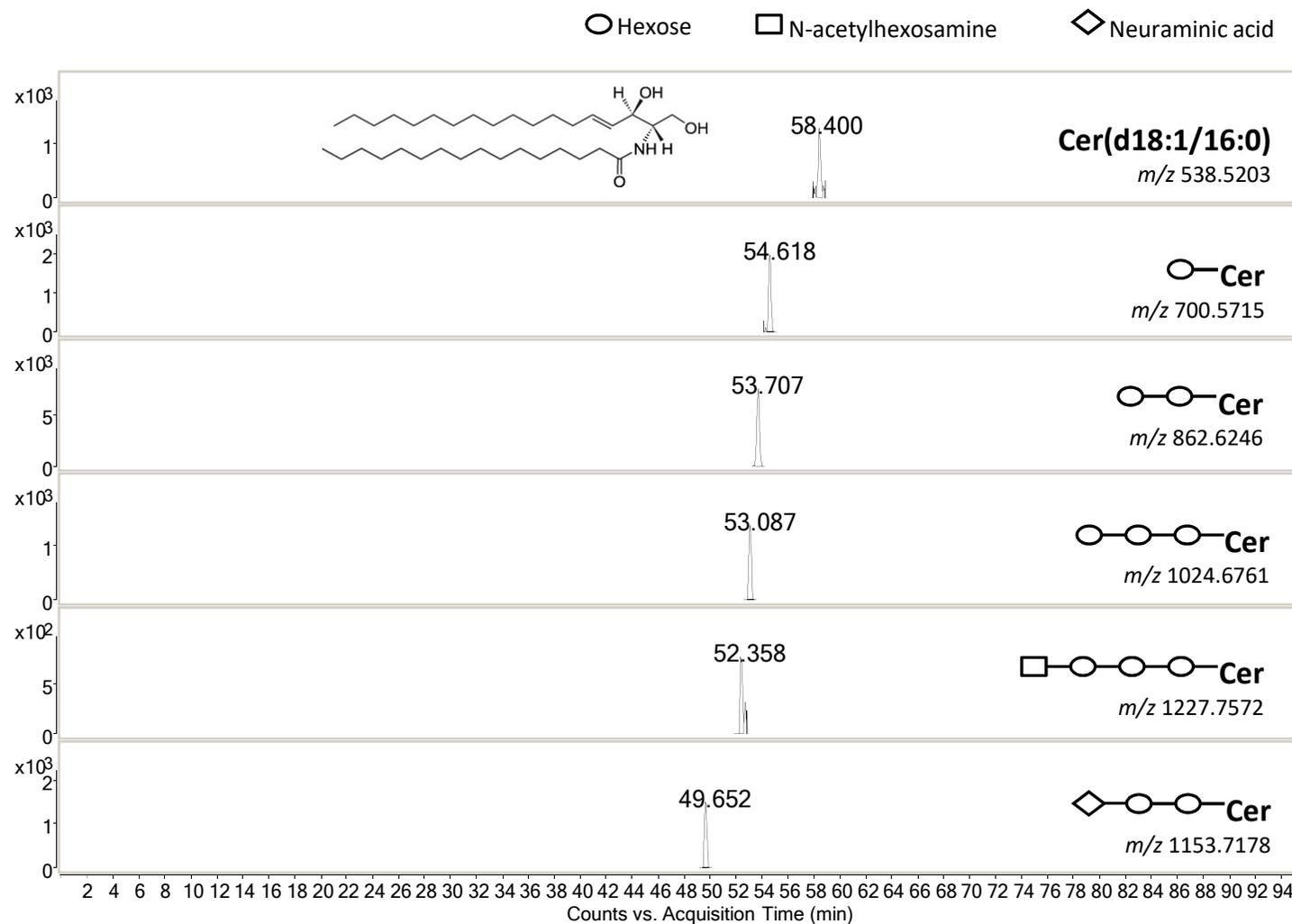
Lipid extract from 100 μl EDTA blood plasma – 50 nl injection  
30 to 98% B in 60 min – 750 nl/min  
A: 20mM NH<sub>4</sub>HCO<sub>2</sub> (PH 5)  
B: C<sub>3</sub>H<sub>7</sub>OH/CH<sub>3</sub>OH (90/10) (v/v)

# μPAC™-MS Lipidomics Platform

## Inter-class separation of ceramide glycosphingolipids



RIC | Research Institute  
for Chromatography



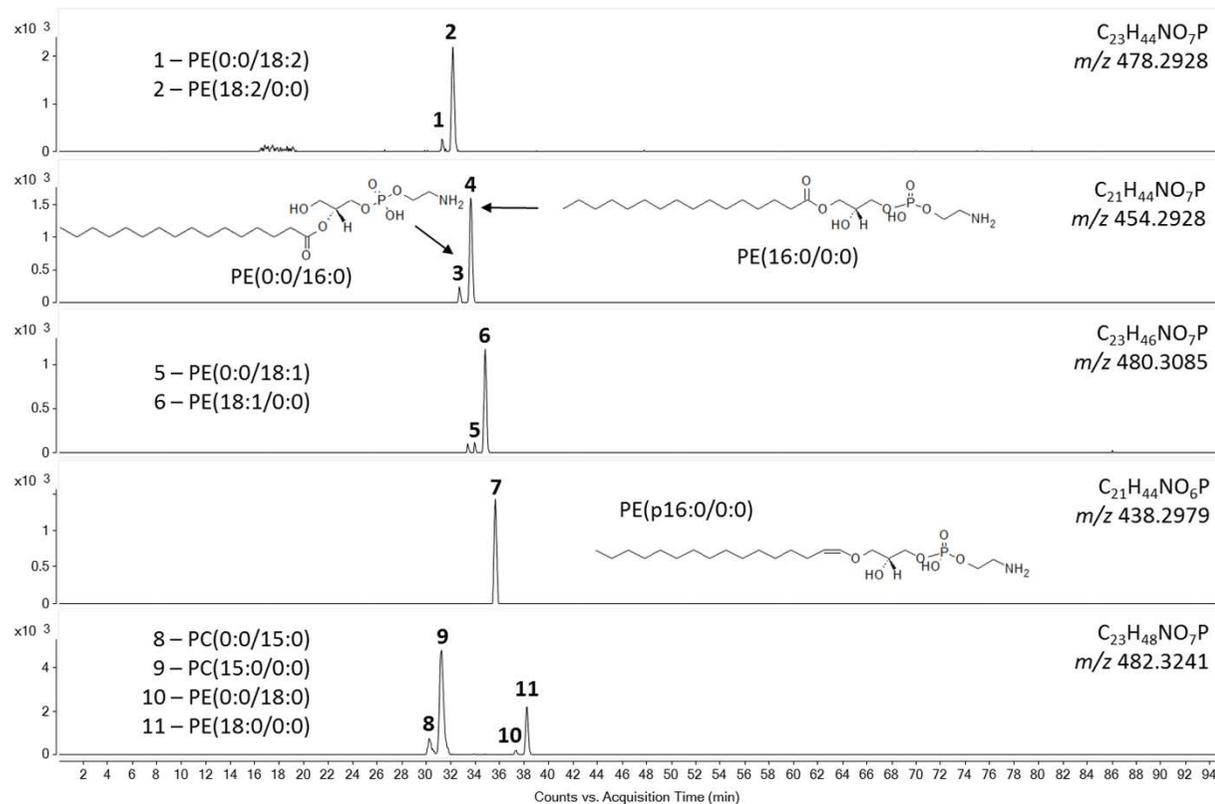
### Separation by:

→ Glycosylation  
pattern

- Polarity increases with number of sugar chains
- More polar molecules elute earlier in RPLC

# $\mu$ PAC<sup>TM</sup>-MS Lipidomics Platform

## Intra-class separation of Lysophosphatidylethanolamines



### Separation by:

#### → Number of carbon atoms

- Lyso-PE (16:0) → peaks 3 & 4
- Lyso-PE (18:0) → peaks 10 & 11

#### → Degree of saturation

- Lyso-PE (18:2) (2 C=C bonds) → peaks 1 & 2
- Lyso-PE (18:1) (1 C=C bond) → peaks 5 & 6
- Lyso-PE (18:0) (no C=C bonds) → peaks 10 & 11

#### → Fatty acid chain position

- Lyso-PE (0:0/16:0) (sn-2 position) → peak 3
- Lyso-PE (16:0/0:0) (sn-1 position) → peak 4

#### → Fatty acid composition

- Lyso-PC (15:0) → peaks 8 & 9
- Lyso-PE (18:0) → peaks 10 & 11

---

# μPAC™-MS Lipidomics Platform

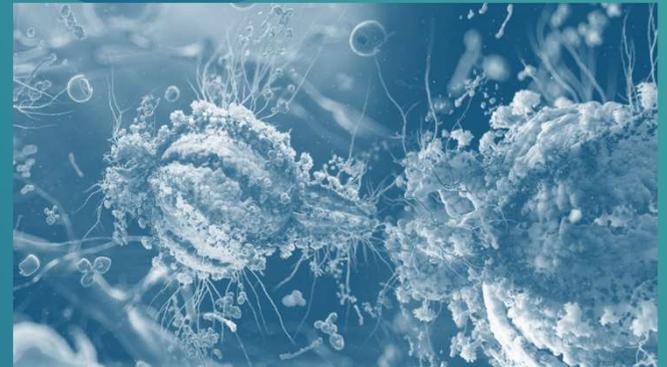
## Conclusions



- The use of the μPAC™ column in lipidomics has been illustrated
- In combination with high resolution mass spectrometry:
  - high lipidome coverage can be obtained
  - inter- and intra-class separation can readily be achieved
  - isomeric lipids can be resolved
- The μPAC™ technology holds great potential in lipidomics studies.

# Data and results

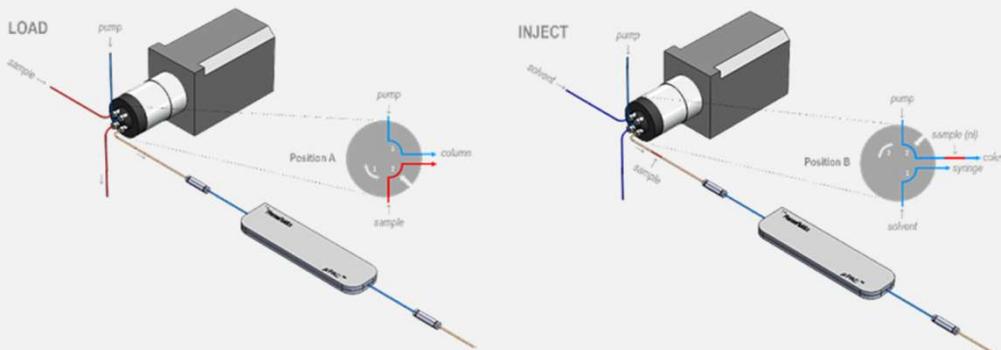
- $\mu$ PAC™-MS Proteomics Platform
  - $\mu$ PAC™-MS Lipidomics Platform
  - **$\mu$ PAC™-MS Metabolomics Platform**
  - $\mu$ PAC™-MS Biopharmaceuticals
- 



# μPAC™-MS Metabolomics Platform

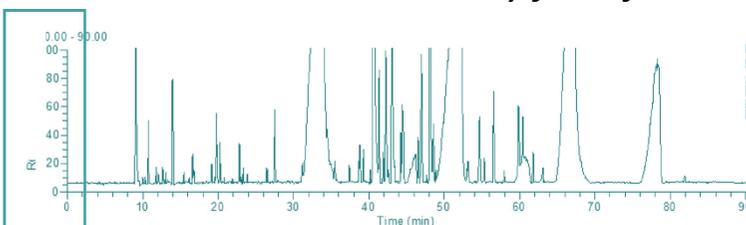


## Plant Metabolomics



*Increased intensity for a factor 1,250 less sample load*

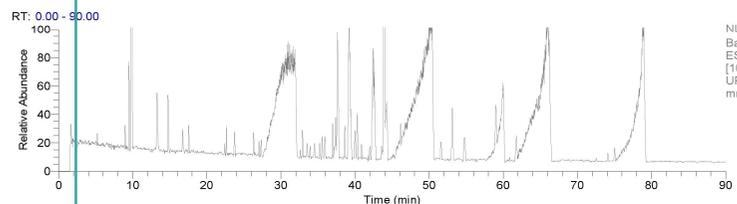
1260 Infinity Nanoflow LC  
Nano ESI (Agilent)  
LTQ-FT Ultra



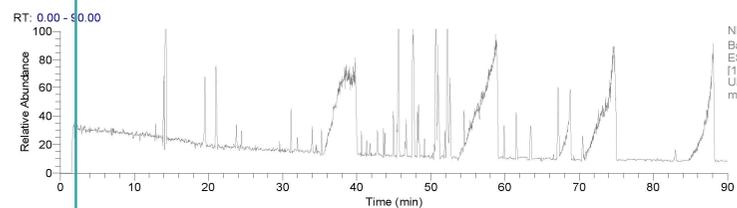
μPAC™ 200 cm; C18

Poplar bark metabolite extract – 4 nl injection  
1 to 50% B in 120 min – **1 μl/min**

Accela (Thermo)  
ESI  
LTQ-FT Ultra



Ref A: C18 (15 cm x 2.1 mm; 1.7 μm particles)  
Poplar bark metabolite extract – 5 μl injection  
1 to 50% B in 120 min – **300 μl/min**



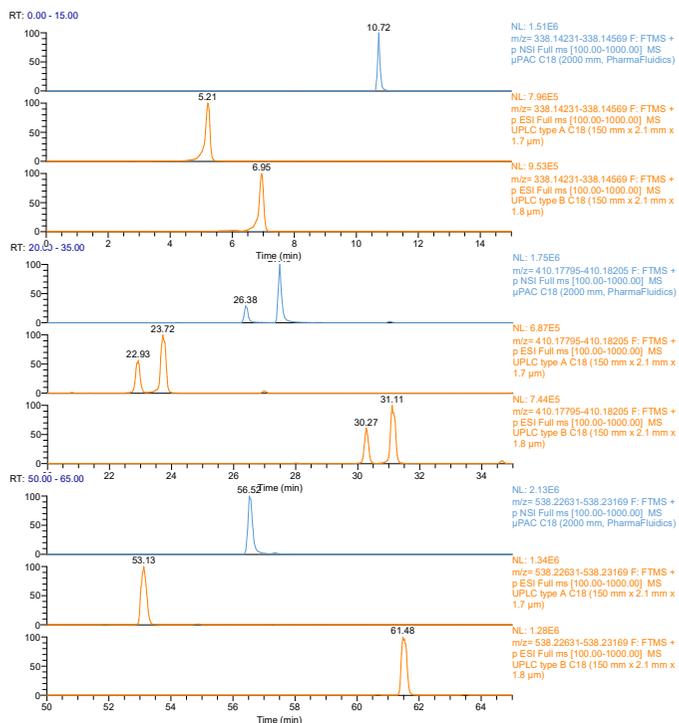
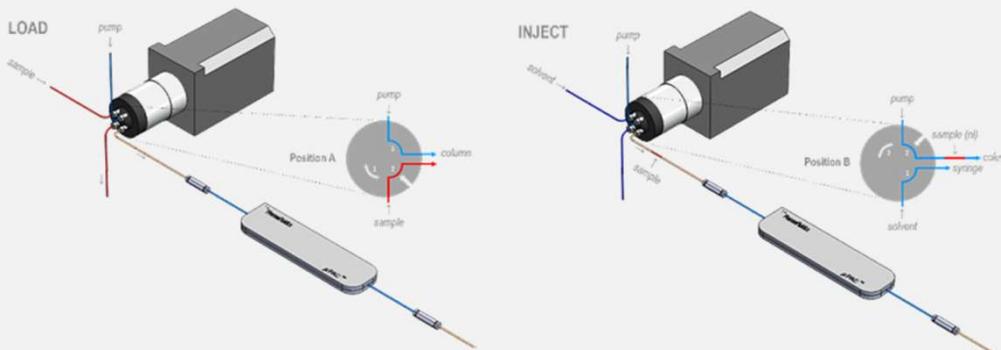
Ref B: C18 (15 cm x 2.1 mm; 1.8 μm particles)  
Poplar bark metabolite extract – 5 μl injection  
1 to 50% B in 120 min – **300 μl/min**

A: H<sub>2</sub>O (100%) with 0,1% FA; B: C<sub>2</sub>H<sub>3</sub>N/H<sub>2</sub>O (80/20) with 0,1% FA

# μPAC™-MS Metabolomics Platform



## Plant Metabolomics



	RT [min]	FWHM [min]	Intensity [I]
<b>PharmaFluidics μPAC™ nano LC C18 (200 cm)</b>	10.72	0.11	1.51E+06
UPLC Type A C18 (15 cm x 2.1 mm x 1.7 μm)	5.21	0.19	7.71E+05
UPLC Type B C18 (15 cm x 2.1 mm x 1.8 μm)	6.95	0.18	9.19E+05
<b>PharmaFluidics μPAC™ nano LC C18 (200 cm)</b>	27.49	0.11	1.74E+06
UPLC Type A C18 (15 cm x 2.1 mm x 1.7 μm)	23.72	0.19	6.61E+05
UPLC Type B C18 (15 cm x 2.1 mm x 1.8 μm)	31.11	0.18	7.25E+05
<b>PharmaFluidics μPAC™ nano LC C18 (200 cm)</b>	56.52	0.15	2.12E+06
UPLC Type A C18 (15 cm x 2.1 mm x 1.7 μm)	53.13	0.21	1.33E+06
UPLC Type B C18 (15 cm x 2.1 mm x 1.8 μm)	61.48	0.20	1.27E+06

# Data and results

- $\mu$ PAC™-MS Proteomics Platform
  - $\mu$ PAC™-MS Lipidomics Platform
  - $\mu$ PAC™-MS Metabolomics Platform
  - **$\mu$ PAC™-MS Biopharmaceuticals**
- 



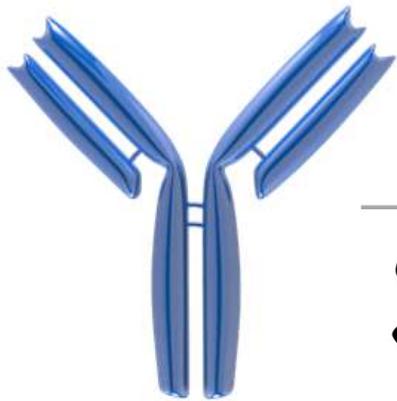
---

# Next-generation mAbs

- The success of mAbs as therapeutics has triggered the development of various next-generation formats such as:
  - **Antibody-drug conjugates (ADC)**
  - Bispecific mAbs
  - Antibody fragments (Fab, Fc, Nanobodies)
  - Fc fusion proteins
  - Glyco-engineered mAbs
- ADCs consist of an antibody that targets a specific tumor cell and a highly potent cytotoxin
- Promise of ADCs: selective delivery of toxic compounds to tumor cells (side effect much lower compared to classical chemotherapy)

# μPAC™-MS Biopharmaceuticals

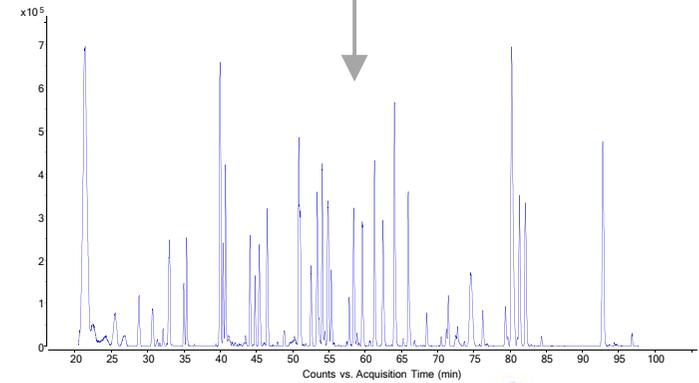
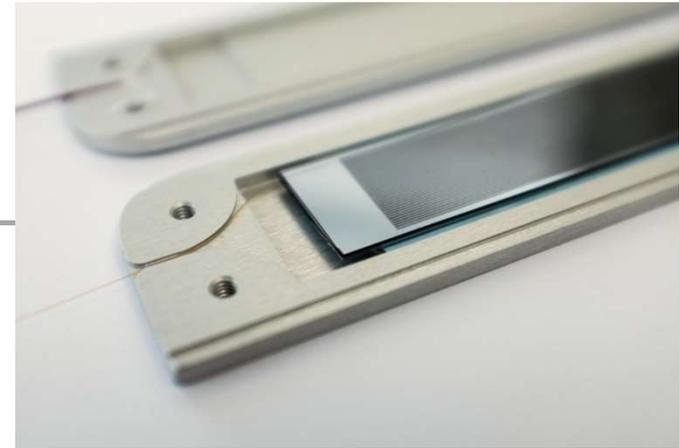
## Workflow



Trypsin

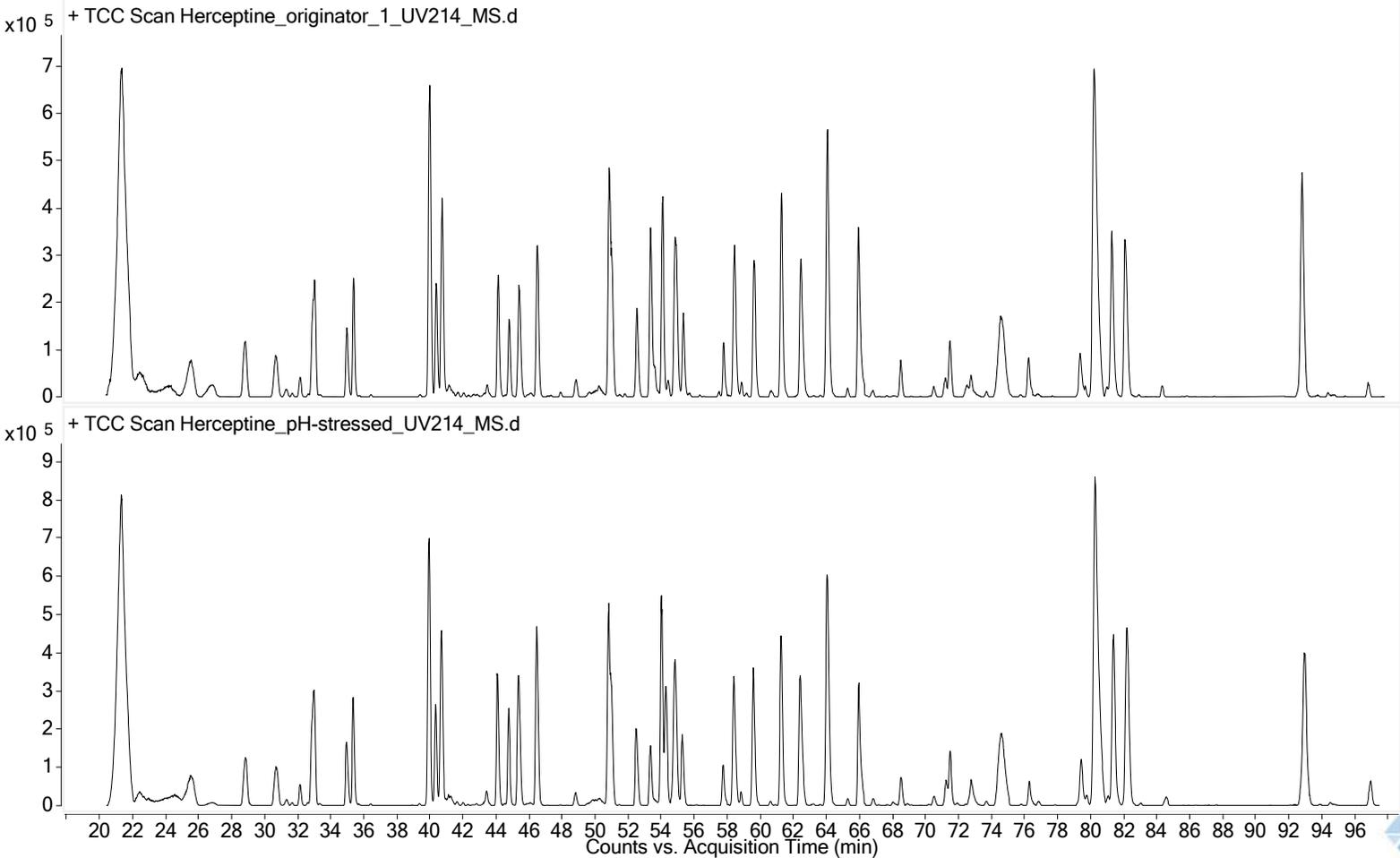


DIQMTQSPSSLSASVGDR  
VTITCR  
ASQDVNTAVAWYQQKPGK  
APK  
LLIYSASFLYSGVPSR  
...



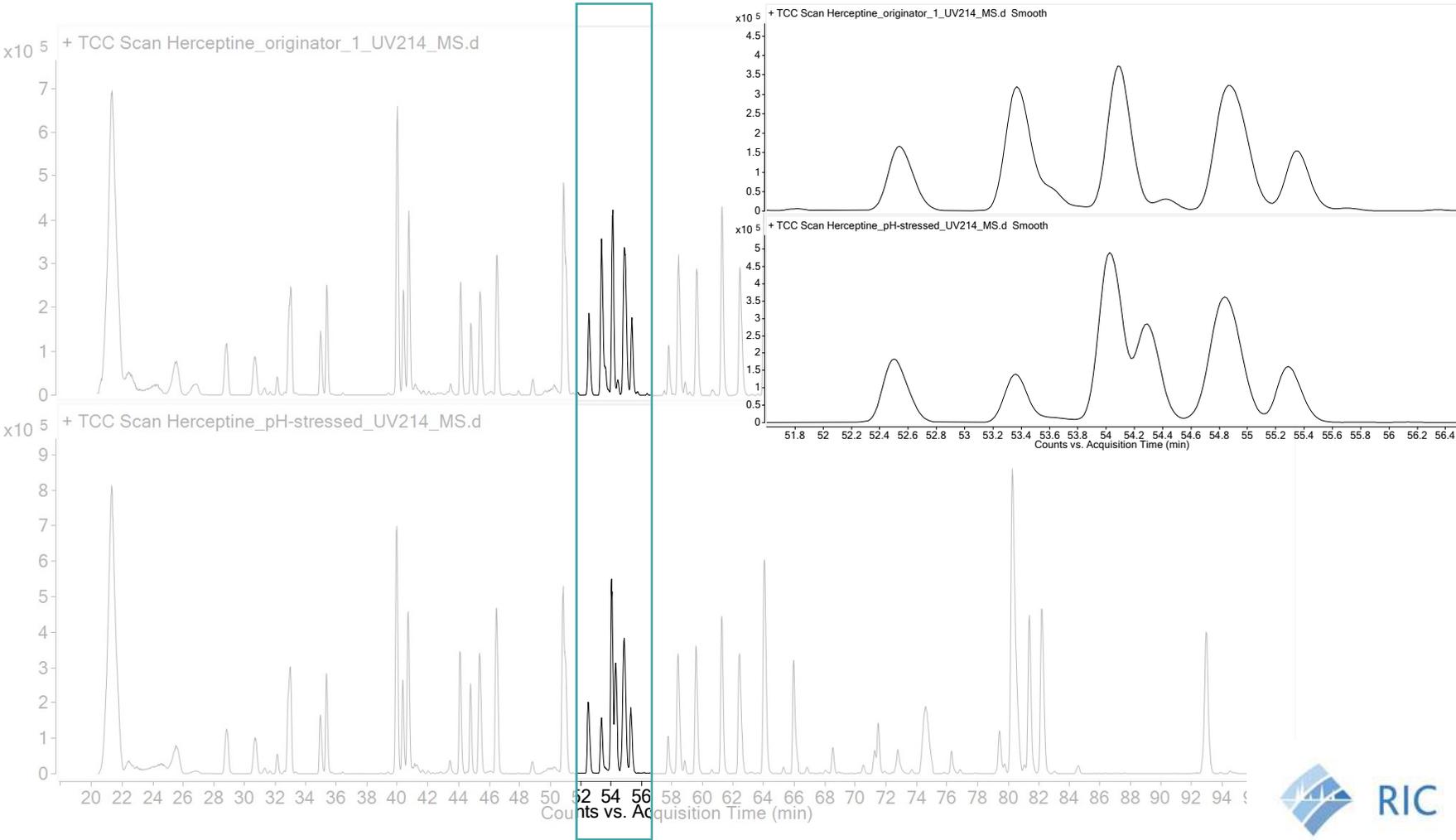
# μPAC™-MS - Biopharmaceuticals

## Herceptin – Originator versus high pH-stressed



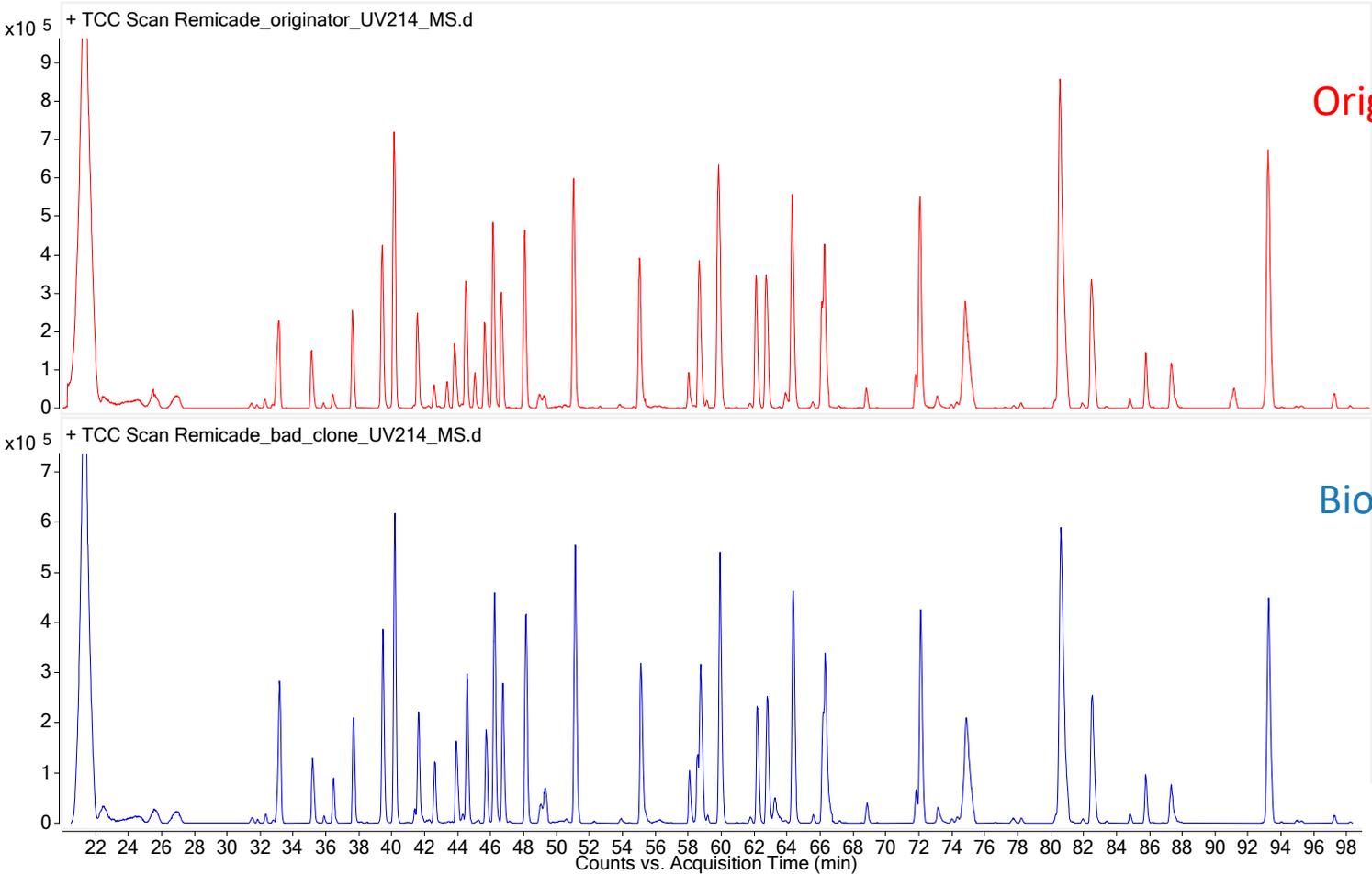
# μPAC™-MS - Biopharmaceuticals

## Herceptin – Originator versus high pH-stressed



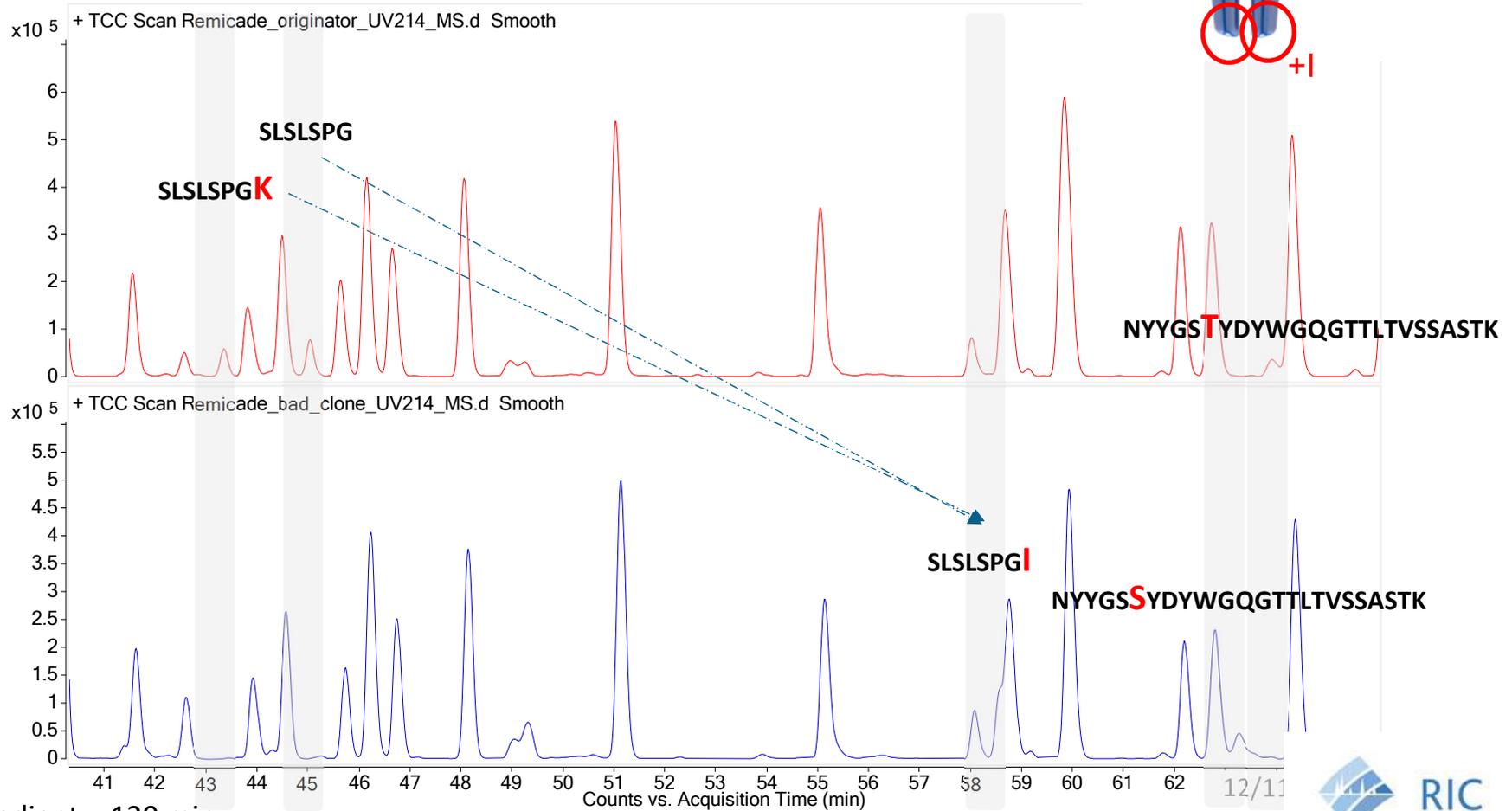
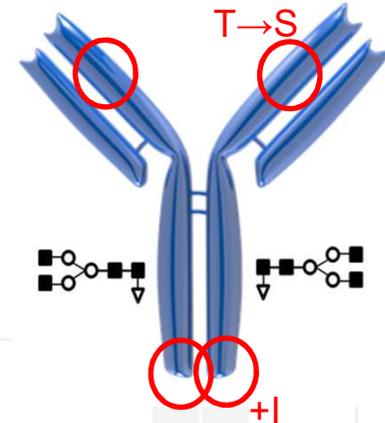
# μPAC™-MS - Biopharmaceuticals

## Remicade - originator vs bad clone



# μPAC™-MS - Biopharmaceuticals

## Remicade - originator vs bad clone



Gradient = 120 min

---

# μPAC™-MS - Biopharmaceuticals



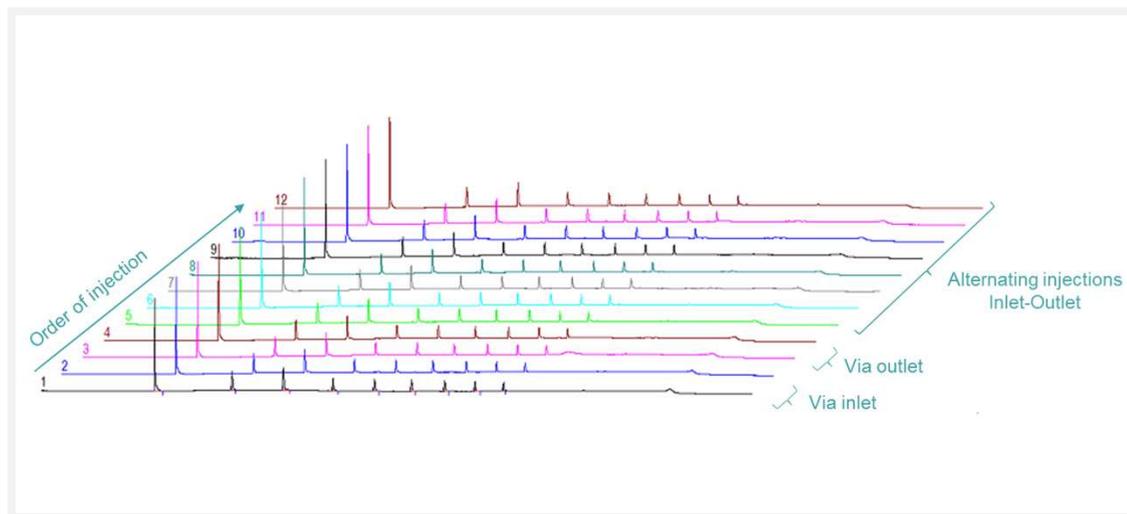
## Conclusions

- The use of the μPAC™ column in the characterization and comparison of mAbs and ADCs has been illustrated.
- In combination with high resolution mass spectrometry, the μPAC™ allows to:
  - confirm the primary structure (sequence)
  - highlight post-translational modifications and amino acid substitutions
  - identify drug conjugations sites
- The μPAC™ column holds great potential in biopharmaceutical applications.

# Micro Pillar Array Column ( $\mu$ PAC™) Robustness

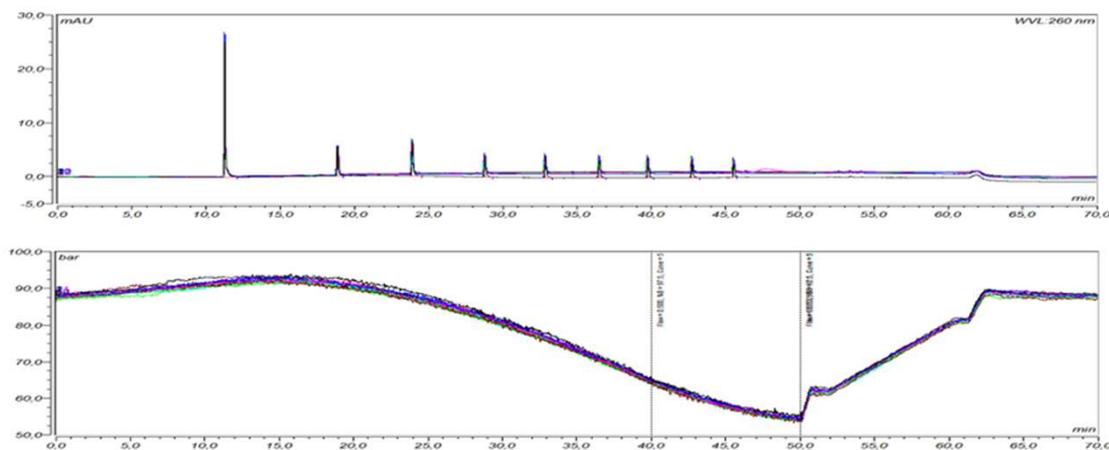
---

# Robustness - Bidirectionality



## Experimental conditions

9 phenones – 4nl injection  
1 to 40% ACN / 0,05% FA  
40 min gradients / 500nl/min



## Conclusions

### Bidirectional use

$\mu$ PAC™ can be used in both straight and reverse directions without loss in performance

# Longevity - Robustness – Acidic pH resistance

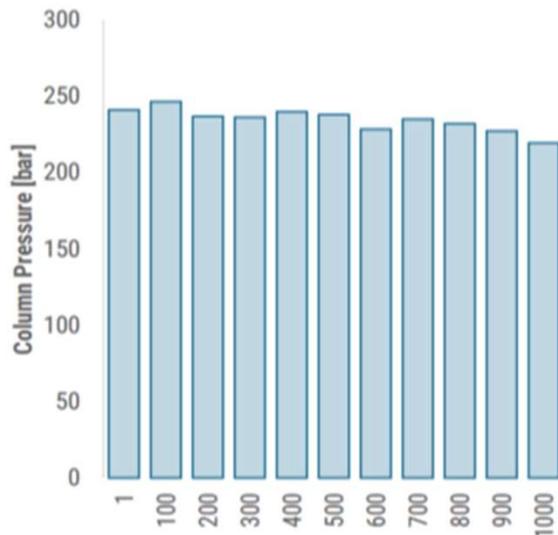
- Number of HeLa cell digest injections
- Total number of injections
- Total volume through column (ml)
- Total amount of column volumes

1000  
3.526  
195,4  
21.707

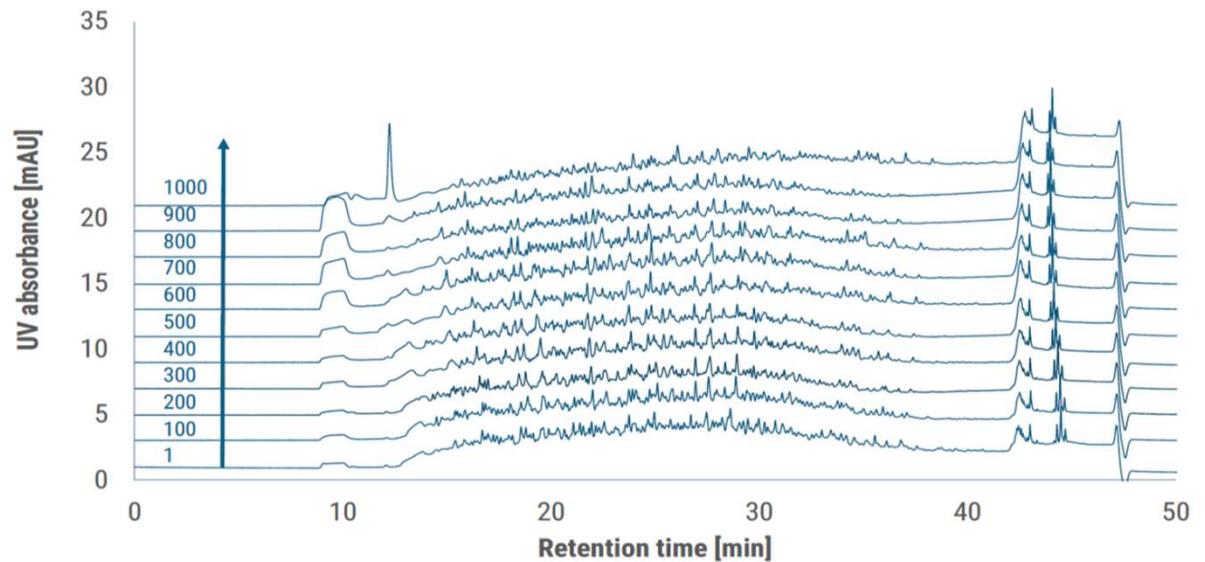
1  $\mu$ l injection  
30 min gradient 1% B  $\rightarrow$  50% B  
0.1% TFA (pH 1,98)  
UV detection: 214nm

100 ng HeLa digest  
Blank  
500 fM Cyto C digest

Pressure profile – HeLa digest



UV profile – HeLa digest



# Longevity - Robustness – Acidic pH resistance

- Number of HeLa cell digest injections
- Total number of injections
- Total volume through column (ml)
- Total amount of column volumes

1000  
3.526  
195,4  
21.707

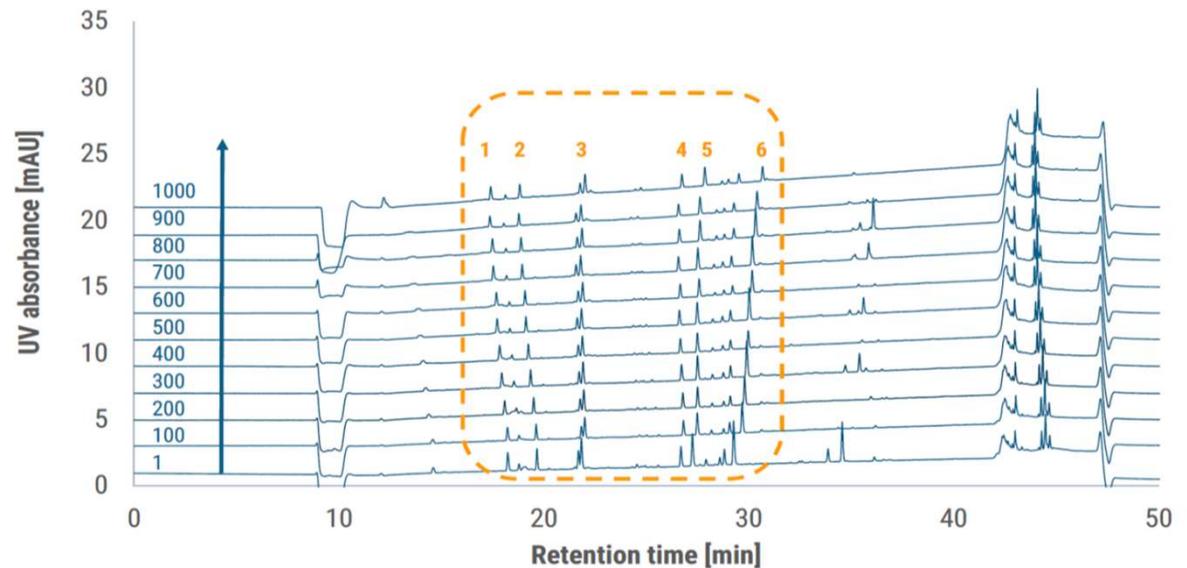
1 µl injection  
30 min gradient 1% B -> 50% B  
0.1% TFA (pH 1,98)  
UV detection: 214nm

100 ng Hela digest  
Blank  
500 fM Cyto C digest

## Chromatographic metrics – Cyto C

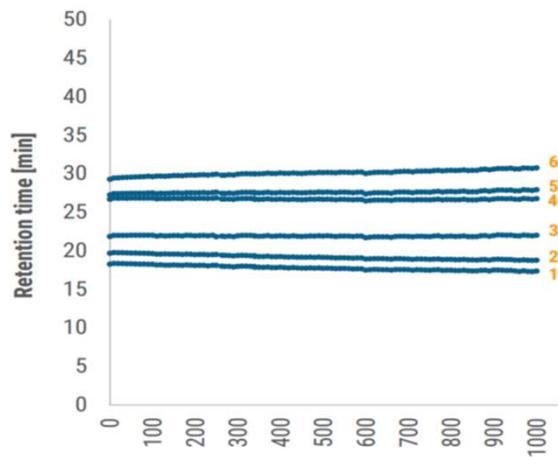
Peptide	Value (min)	Std DEV	CV (%)
1	17.76	0.30	1.72
2	19.17	0.31	1.59
3	21.91	0.07	0.32
4	26.70	0.08	0.31
5	27.59	0.13	0.48
6	30.12	0.35	1.15
Average PW	0.14		

## UV profile – Cyto C digest

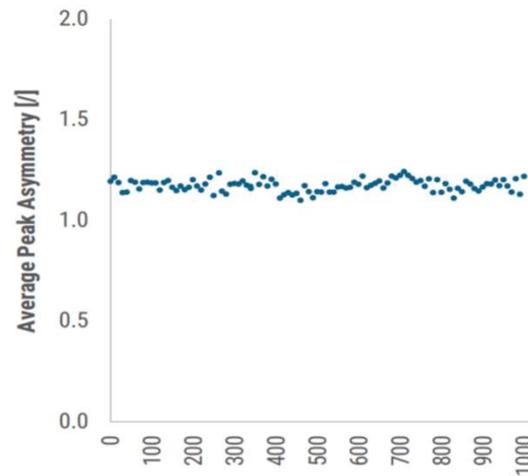


# Longevity - Robustness – Acidic pH resistance

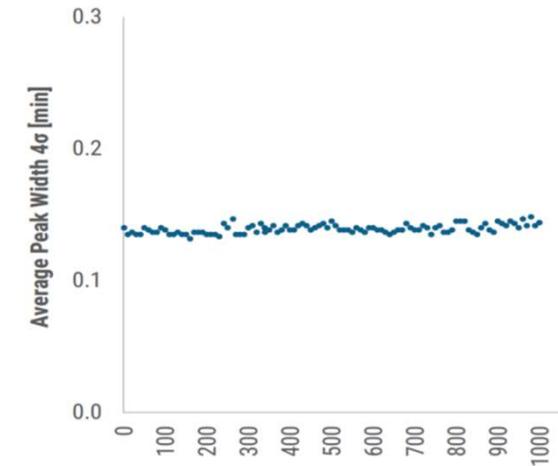
Retention



Peak asymmetry



Peak width



## Conclusion

The  $\mu$ PAC™ can be used up to 1000 injections of commercial HeLa cell digest (> 3500 total injections) in a low pH range without any loss of performance or pressure increase.

# Robustness - Longevity – Challenging samples

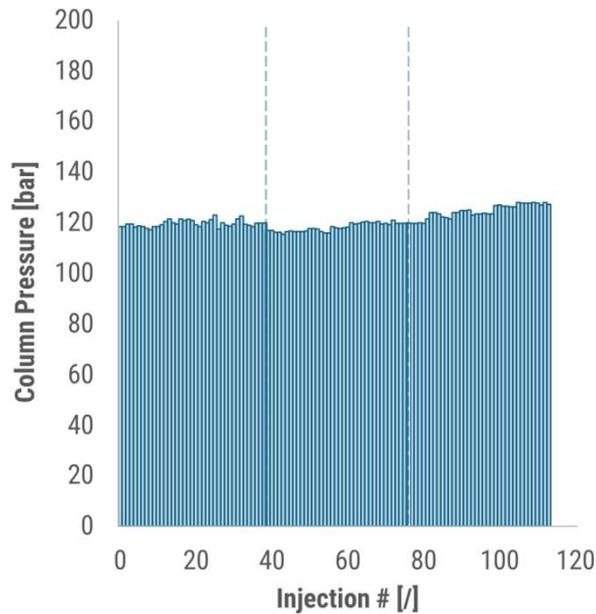
3 different sets of "challenging" samples (n=6):

Set 1: Human cell line treated with detergent (Triton X-114)

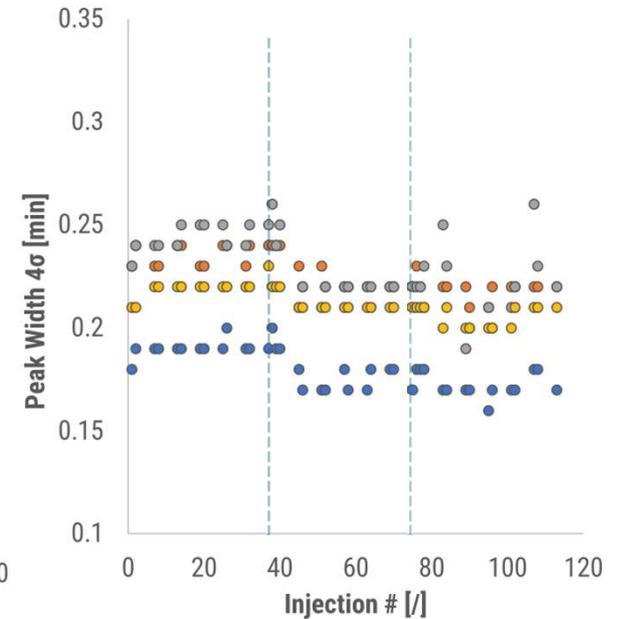
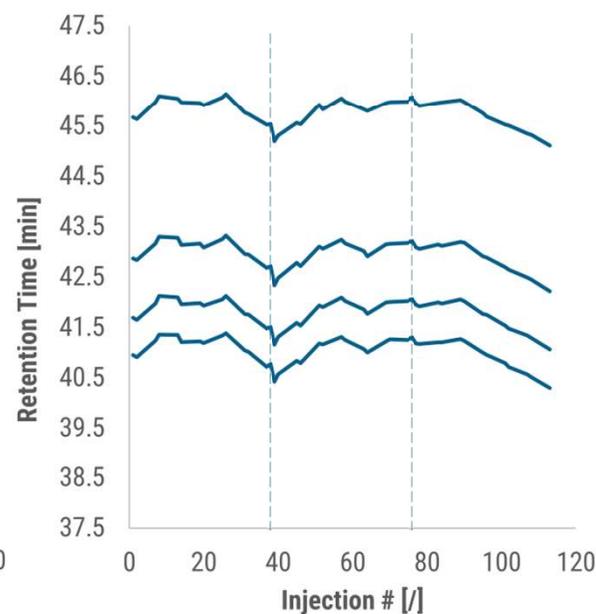
Set 2: Plant material lysed with detergent (NP-40)

Set 3: TnT quick coupled transcription/translation system with black precipitation

All injections



Cytochrome C digest reference peptides



## Conclusion

No significant increase in column backpressure and excellent RT stability (CV < 1%) was observed.

# Micro Pillar Array Column ( $\mu$ PAC™) RT Stability

---

# RT stability – PRTC mixture

## Experimental conditions:

Pierce RT calibration mixture (P/N 88320) – 50 fmoles spiked in 100 ng HeLa digest  
 1 to 40% ACN / 0,1% FA  
 30 min gradient / 750 nl/min

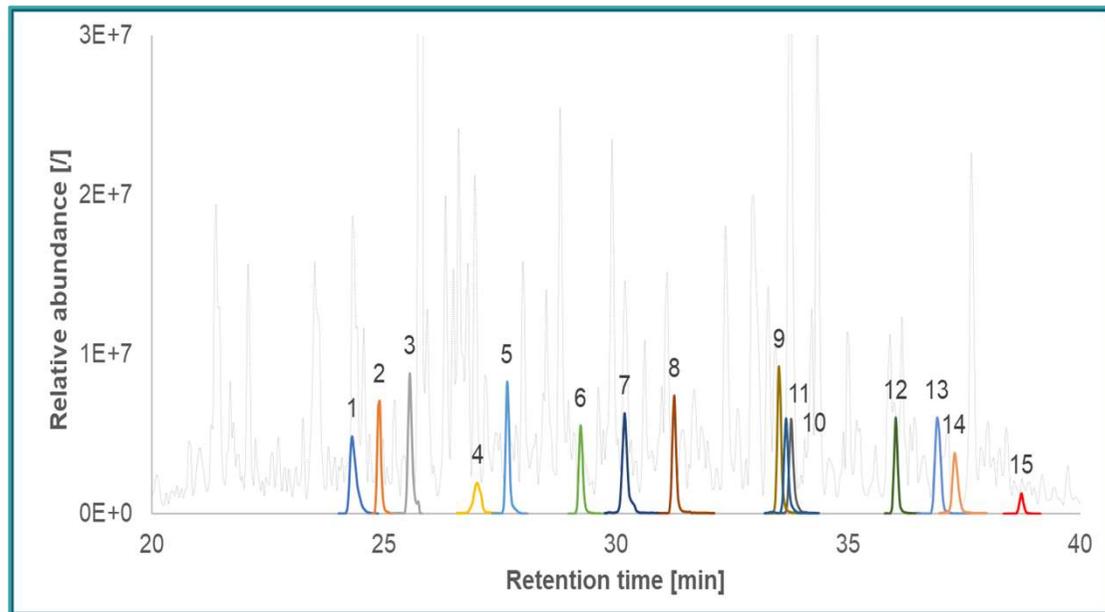
## Instrumental setup:

Thermo Scientific™ Ultimate 3000 RSLC  
 1 µL direct injection  
 200 cm µPAC column @ 30°C  
 Thermo Scientific™ LTQ Orbitrap XL™

## Chromatographic metrics (n=3)

Peptide	Sequence	m/z	RT [min]	
			Value	% CV
1	SSAAPPPPPR	493.768	23.98	1.27
2	GISNEGQNASIK	613.317	24.58	1.28
3	HVLTSIGEK	496.287	25.23	1.20
4	DIPVPPKPK	451.283	26.69	1.17
5	IGDYAGIK	422.736	27.34	1.08
6	TASEFSAIAQDK	695.832	28.93	1.02
7	SAAGAFGPESLR	586.8	29.87	1.04
8	ELGQSGVDTYLQTK	773.896	30.92	1.06
9	GLILVGGYGTR	558.326	33.18	0.91
10	GILFVSGVSGGEEGAR	801.412	33.45	0.89
11	SFANQPLEVVYSK	745.392	33.33	0.91
12	LTILEELR	498.802	35.73	0.83
13	NGFILDGFPR	573.303	36.64	0.74
14	ELASGLSFPVGFK	680.374	37.01	0.72
15	LSSEAPALFQFDLK	787.421	38.42	0.79
<b>Mean CV:</b>			<b>0.99</b>	

## Elution profiles – base peak chromatogram



# RT stability – PRTC mixture

## Experimental conditions:

Pierce RT calibration mixture (P/N 88320) – 50 fmoles spiked in 100 ng HeLa digest  
 1 to 40% ACN / 0,1% FA  
 240 min gradient / 300 nl/min

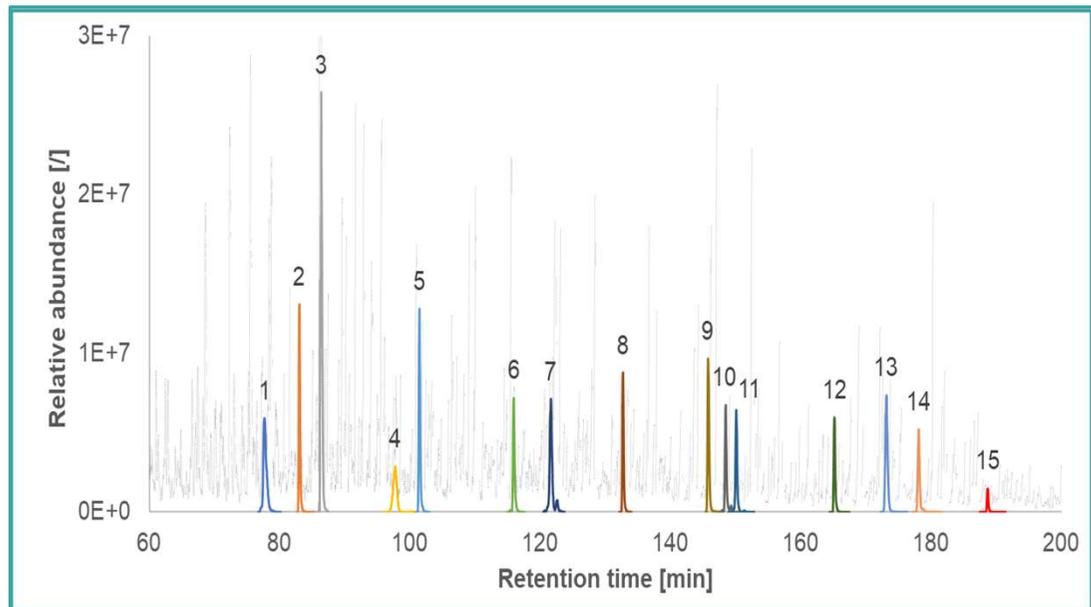
## Instrumental setup:

Thermo Scientific™ Ultimate 3000 RSLC  
 1 µL direct injection  
 200 cm µPAC column @ 30°C  
 Thermo Scientific™ LTQ Orbitrap XL™

## Chromatographic metrics (n=3)

Peptide	Sequence	m/z	RT [min]	
			Value	% CV
1	SSAAPPPPPR	493.768	77.87	0.69
2	GISNEGQNASIK	613.317	83.17	0.61
3	HVLTSIGEK	496.287	86.52	0.64
4	DIPVPKPK	451.283	97.48	0.82
5	IGDYAGIK	422.736	101.16	0.86
6	TASEFSAIAQDK	695.832	115.38	0.87
7	SAAGAFGPESLR	586.8	121.06	0.84
8	ELGQSGVDTYLQTK	773.896	132.07	0.78
9	GLILVGGYGTR	558.326	145.01	0.77
10	GILFVGSVSGGEEGAR	801.412	147.73	0.78
11	SFANQPLEVVYSK	745.392	149.25	0.75
12	LTILEELR	498.802	164.26	0.80
13	NGFILDGFPR	573.303	172.08	0.80
14	ELASGLSFPVGFK	680.374	177.03	0.77
15	LSSEAPALFQFDLK	787.421	187.60	0.75
			<b>Mean CV:</b>	<b>0.77</b>

## Elution profiles – base peak chromatogram



## Conclusion

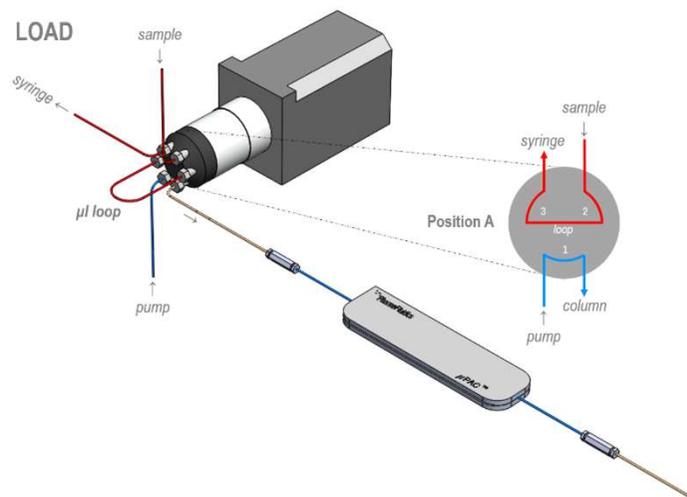
Excellent RT stability was observed with average CV values <1.0% for 750 nL/min and <0.8% for 300 nL/min.

# Connectivity

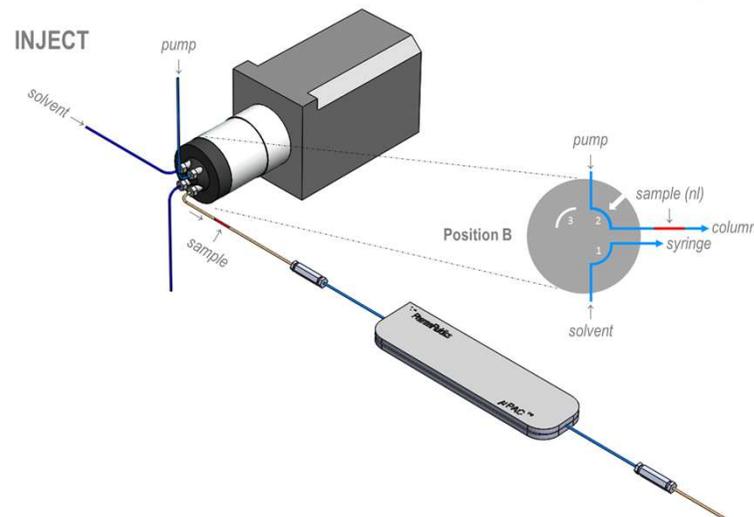
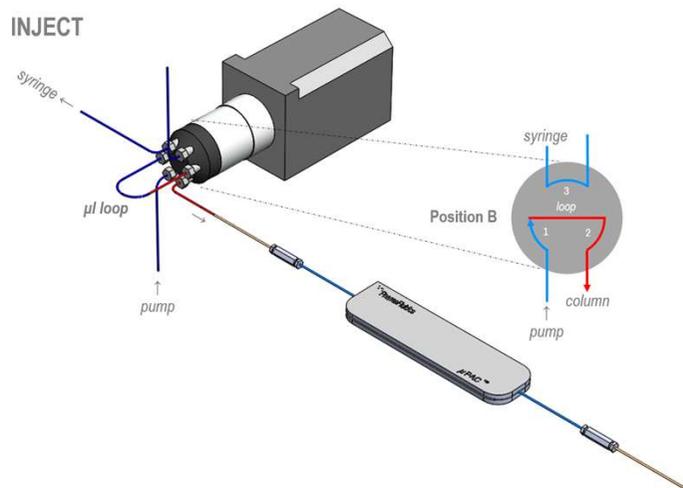
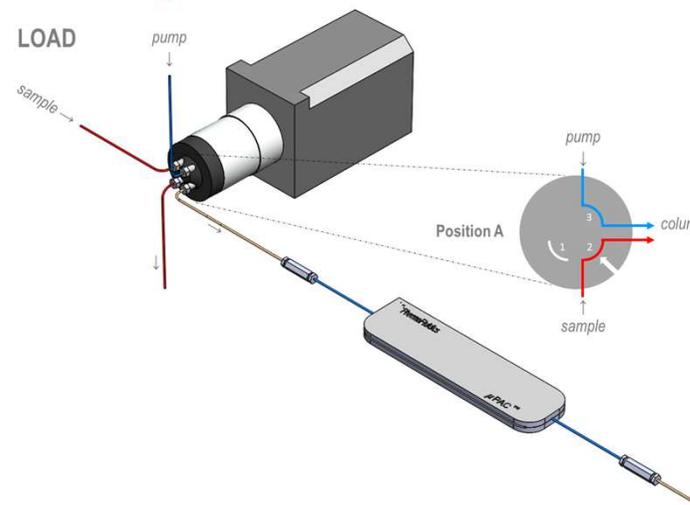
---

# $\mu$ PAC™ Connectivity - Injection

## Direct injection method – External loop



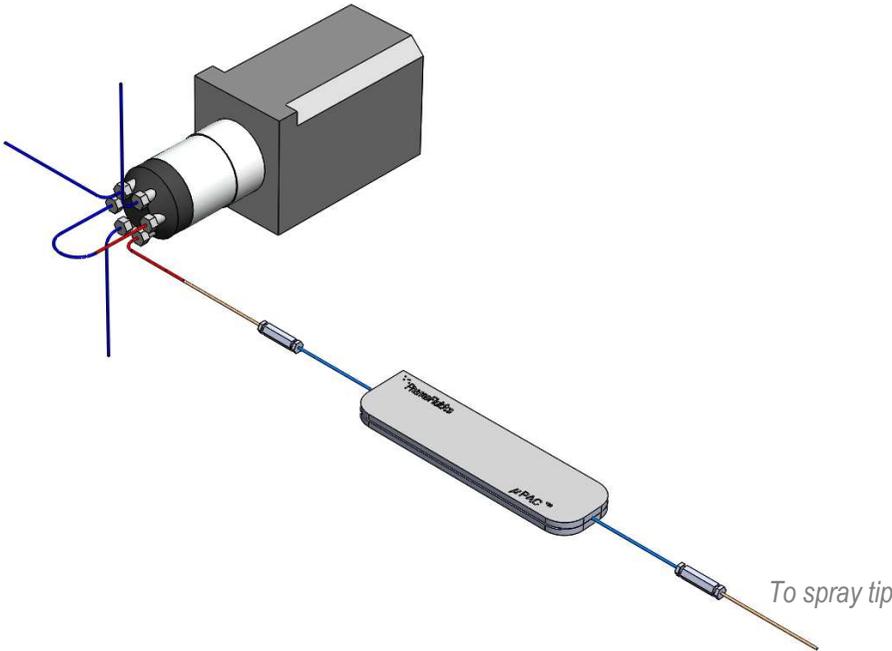
## Direct injection method – Internal loop



[support@pharmafluidics.com](mailto:support@pharmafluidics.com)

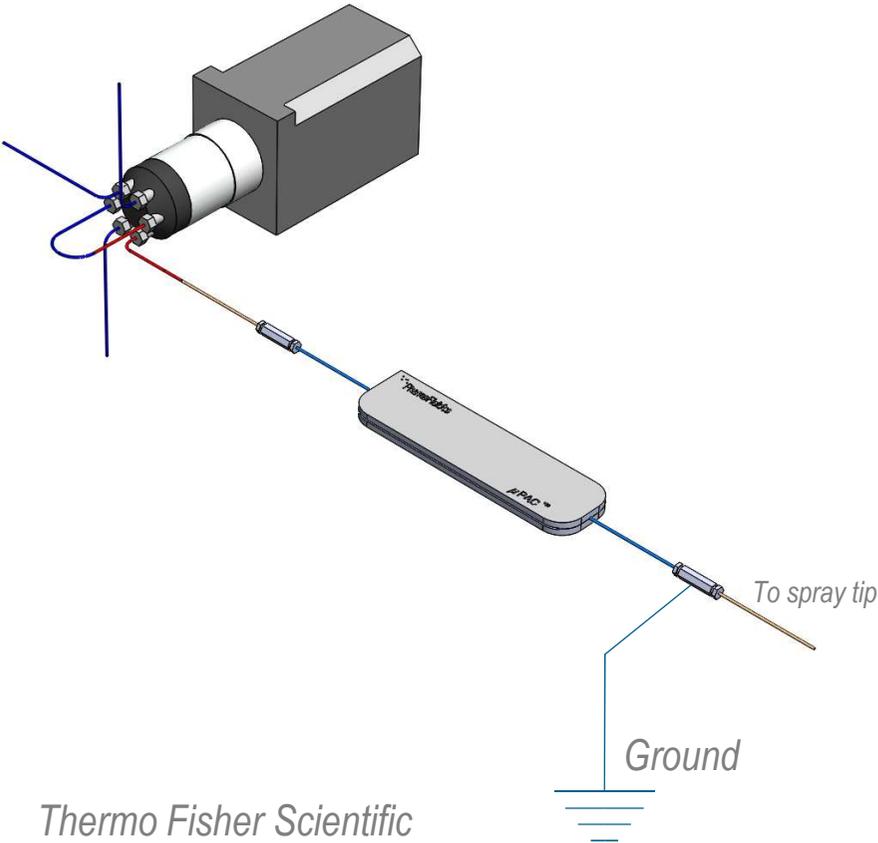
# μPAC™ Connectivity – MS Interfacing

## Grounded spray tip systems



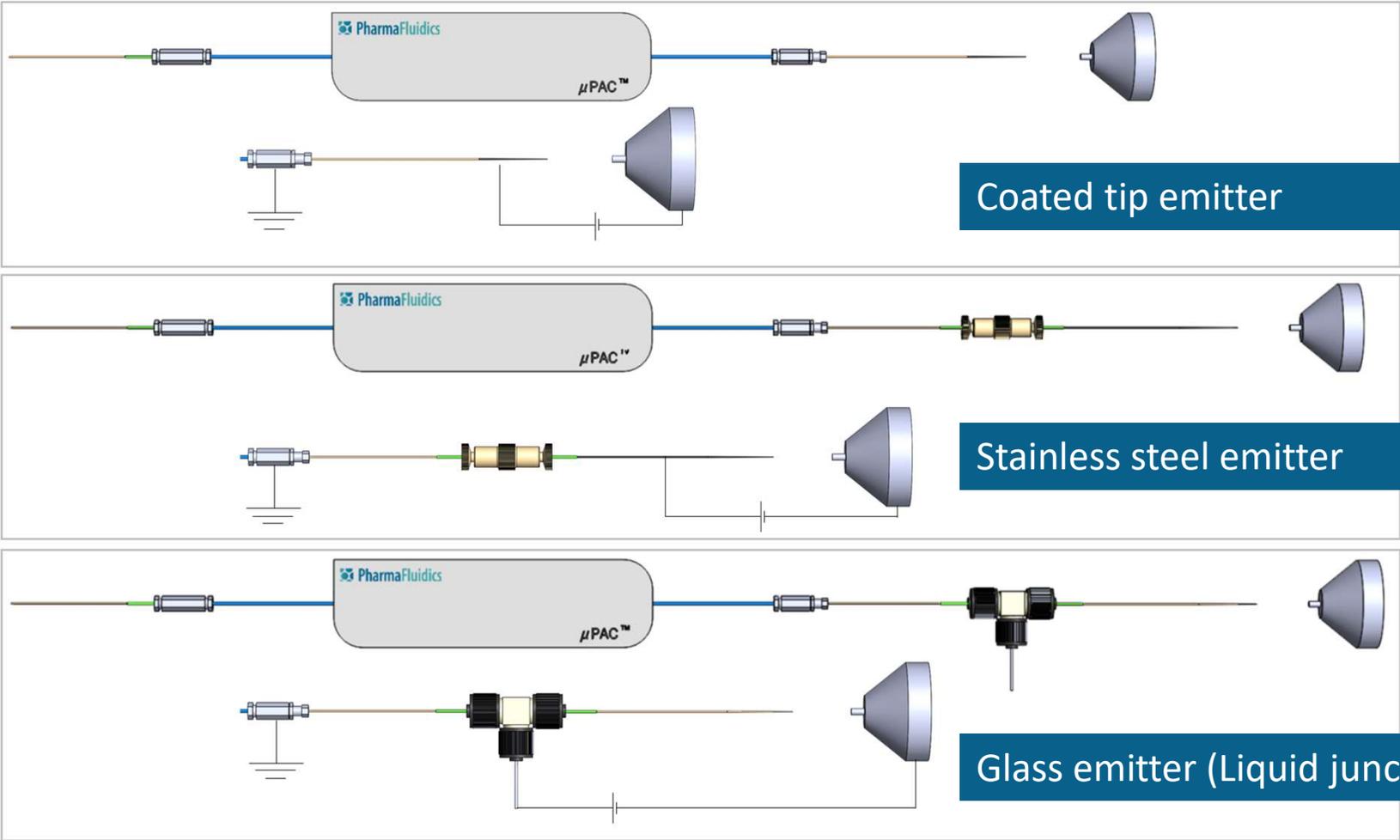
Agilent  
Bruker  
Sciex

## HV spray tip systems

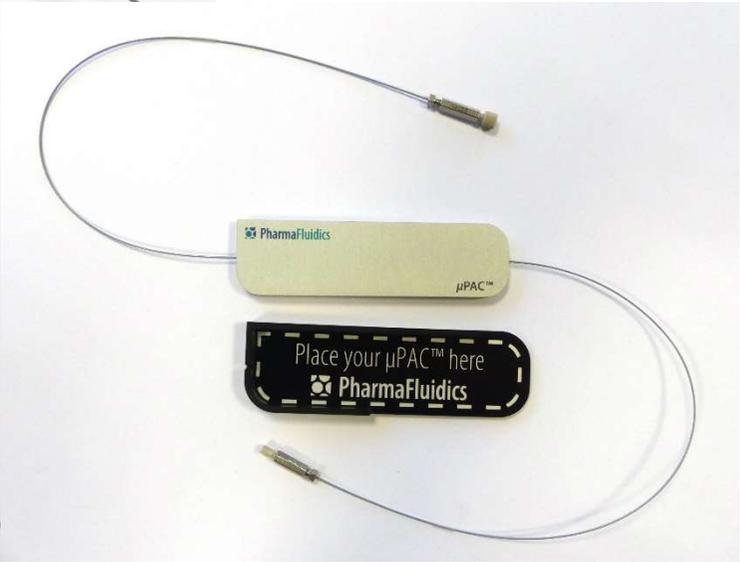
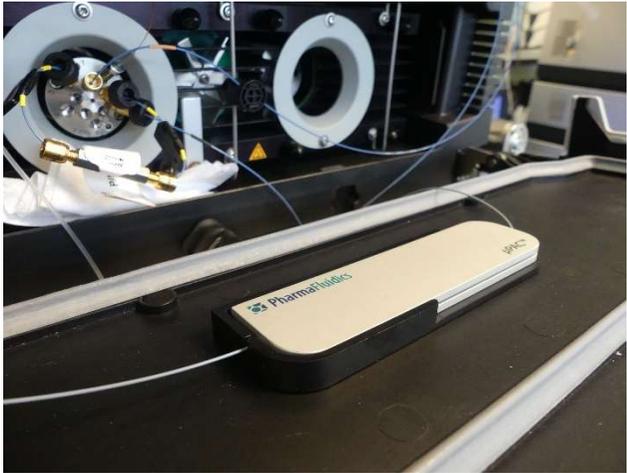
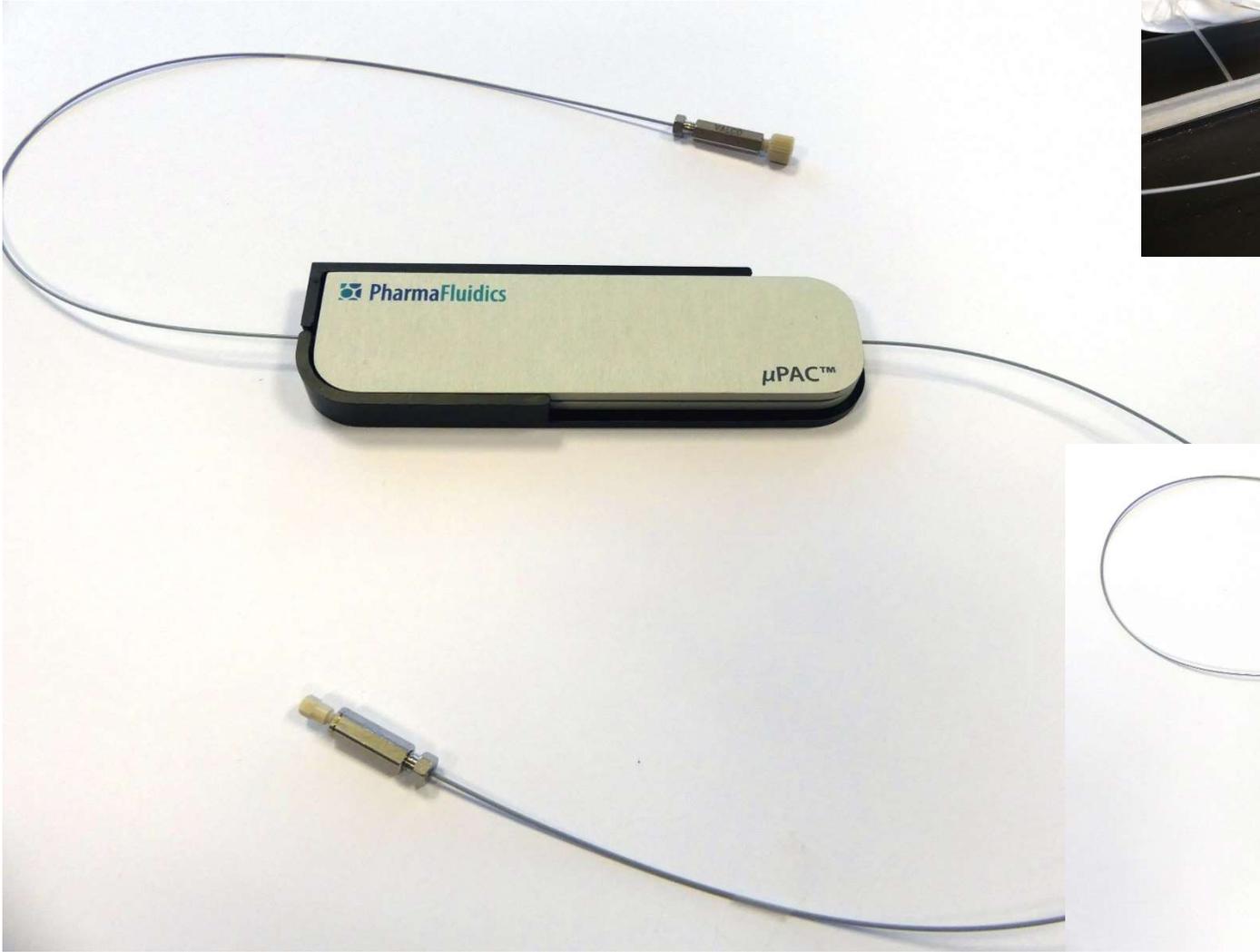


Thermo Fisher Scientific  
Waters

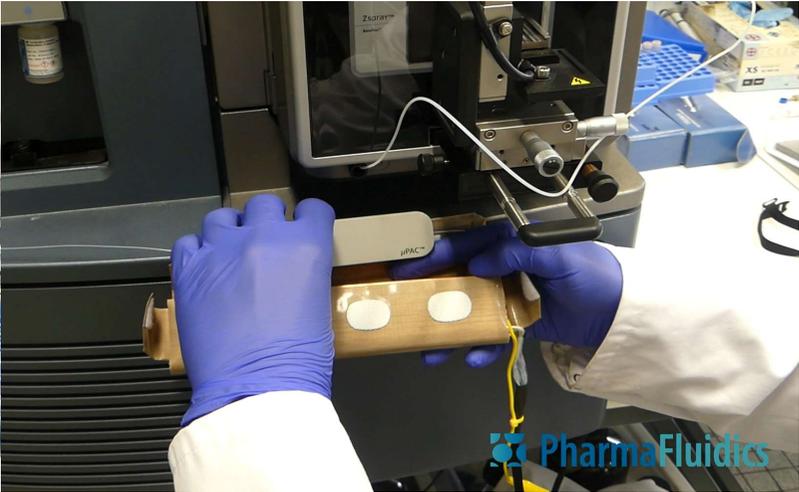
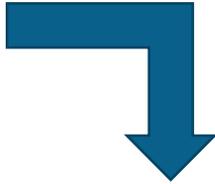
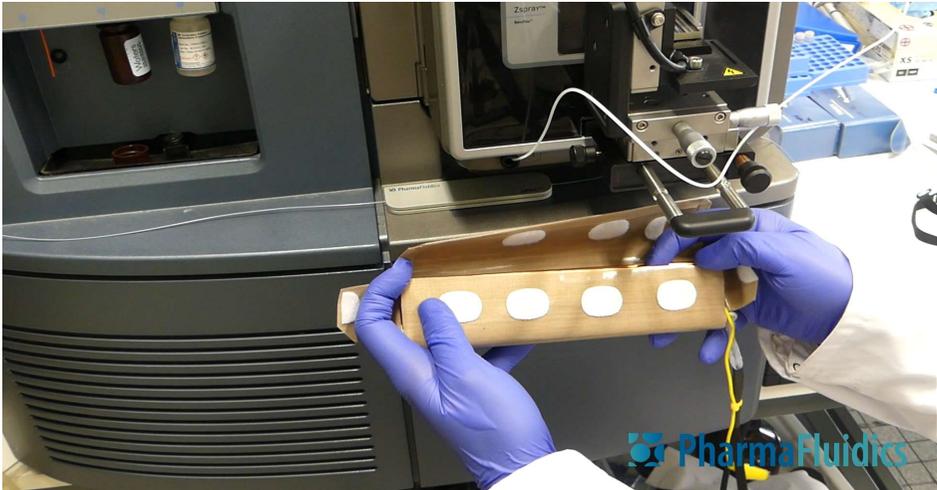
# Connectivity Esi-MS interfacing



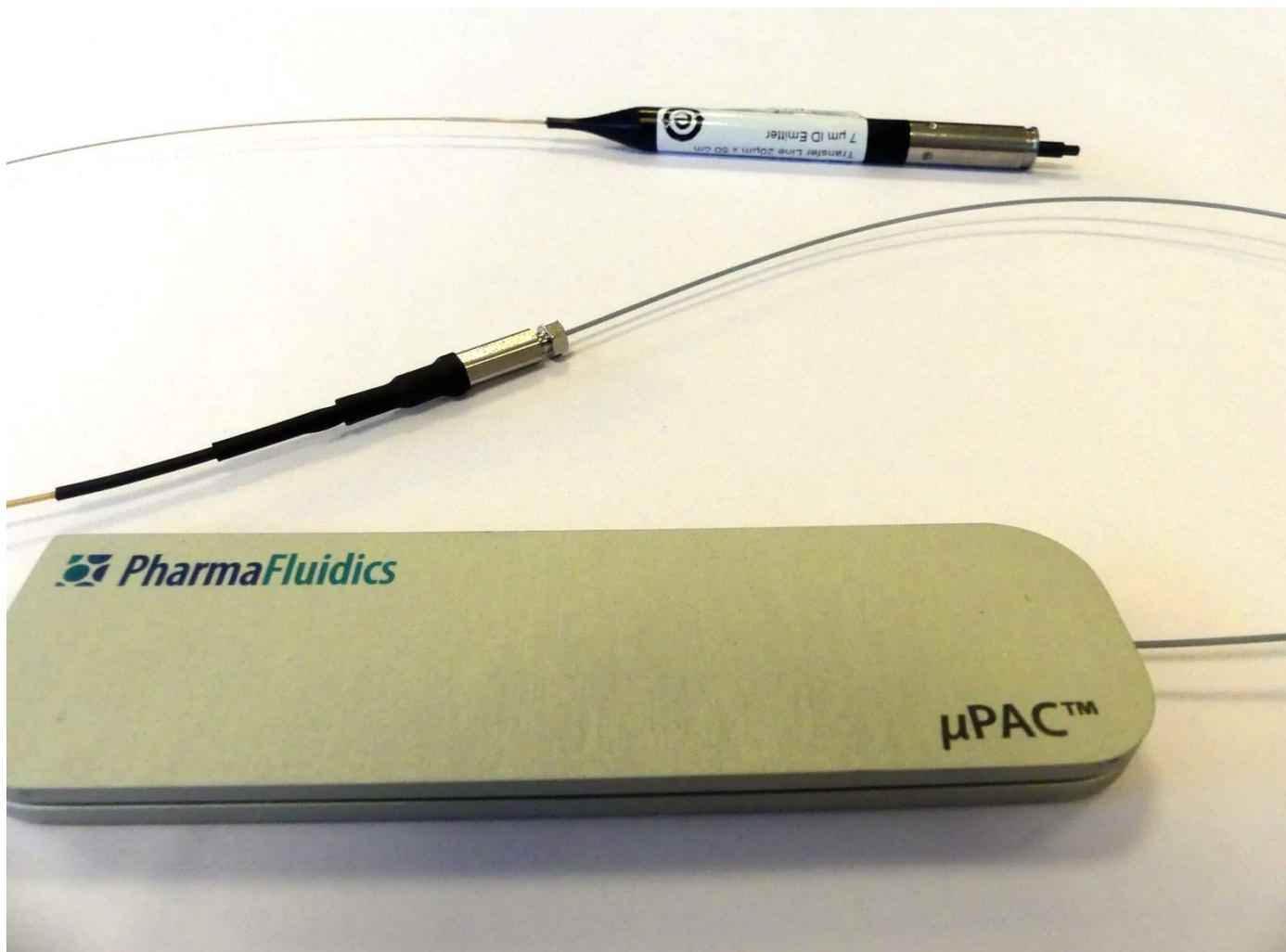
# μPATCH



# Butterfly heater



# $\mu$ PAC™ Compatible EASY-Spray™ Emitter



Do you work with short gradients?

Do you work with a 15cm packed bed?

Do you work with long gradients?

Do you work with a 50cm packed bed?

Are you working in a routine lab?

Are you working with comprehensive proteomics?

Then our 50cm  $\mu$ PAC™ is perfect for you!

- 0.1 – 2  $\mu$ l/min
- 30 min – 2h gradient time
- 30% increase in prot ID

Then the 200cm  $\mu$ PAC™ is your best choice!

- 0.1 – 1  $\mu$ l/min
- 4h– 8h gradient time
- Unique proteome coverage

---

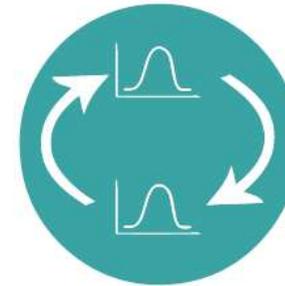
# Why should you use $\mu$ PAC™?



Increased sensitivity  
Due to high degree of order



Unrivalled robustness and  
number of injections  
Due to the solid silicon backbone



Outperforming reproducibility  
Due to the mask-based etching process



Less column replacements  
Due to the high number of injections



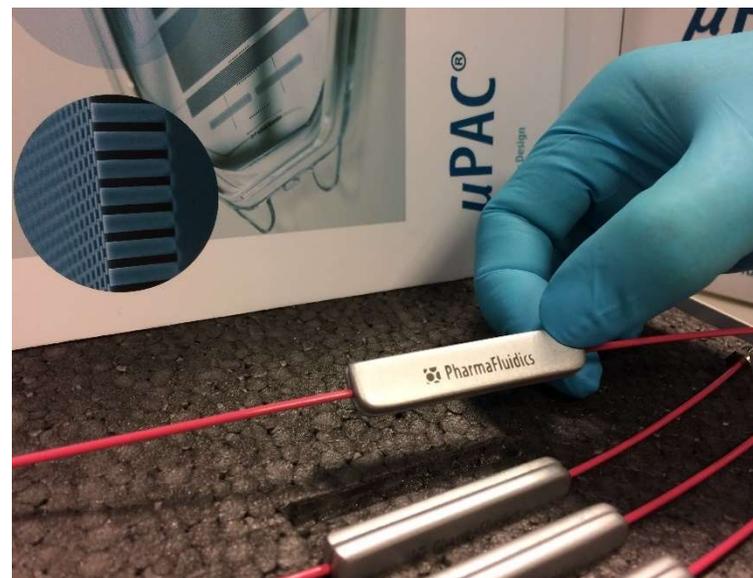
Less prone to clogging  
Due to high column permeability



Sharper peaks and  
excellent separation power  
Due to exceptional column length

# New $\mu$ PAC™ trapping column

- Reducing sample loading times to a minimum with significant higher flow rates
- Instant pressurization due to separation bed of perfectly ordered, free-standing pillars
- Perfectly symmetrical and fritless column design, allowing bidirectional use
- Compatible with both switching valve (regular and backflush elution) or vented trapping configuration



# Order & service enquiries

**Address:**

Technologiepark Zwijnaarde 3  
Gent (Zwijnaarde)  
B-9052, BELGIUM

**Telephone:**

+32 (0)9 241 56 57

[www.pharmafluidics.com](http://www.pharmafluidics.com)

[web.orders@pharmafluidics.com](mailto:web.orders@pharmafluidics.com)

[web.enquiries@pharmafluidics.com](mailto:web.enquiries@pharmafluidics.com)



To serve your custom separation needs,  $\mu$ PAC<sup>®</sup> cartridges are available in various lengths and equipped with connectors matching your application. Stationary phase chemistry is currently focusing on reversed phase HPLC separations.

Product	Stationary phase	Product number
50 cm $\mu$ PAC <sup>™</sup> column	C18 endcapped	5525031518200
200 cm $\mu$ PAC <sup>™</sup> column	C18 endcapped	552503151850
$\mu$ PAC <sup>™</sup> Compatible EASY-Spray <sup>™</sup> Emitter	NA	PhF80000530



MD Scientific is an authorized distributor in Denmark  
[www.md-scientific.dk](http://www.md-scientific.dk) - +45 7027 8565

